

Using Precision Therapies to Protect the Developing Brain from Viral Infection and Injury

Clara Abdelmalek,^{1,2} Ademide Adeyemo,^{1,2} Aubrey Alexander,^{1,2} Zofia Dyba,^{1,2} Caroline Hobson,^{1,2} Haider Hussain,^{1,2} Erin Li,^{1,2} Soumya Maturi,^{1,2} Arnav Patel,^{1,2} Nandi Patel,^{1,2} Ashley Pocasangre,² Hannah Rasmussen,^{1,2} Janet Ruan,^{1,2} Eunbin Park,¹ BS, MS, Erin Tran,² PhD, Youssef Kousa,^{1,2} MS, DO, PhD

¹Center for Genetic Medicine, Children’s National Research Institute
²Gemstone Honors College, University of Maryland



Background

Prenatal viral infections can cause severe damage to the developing brain. Among the most threatening prenatal pathogens, Zika virus can cause microcephaly, ocular abnormalities, congenital contractures, seizures, and fetal demise, termed congenital Zika syndrome.¹ Currently, there is no prenatal standard of care or treatment for Zika infection.²

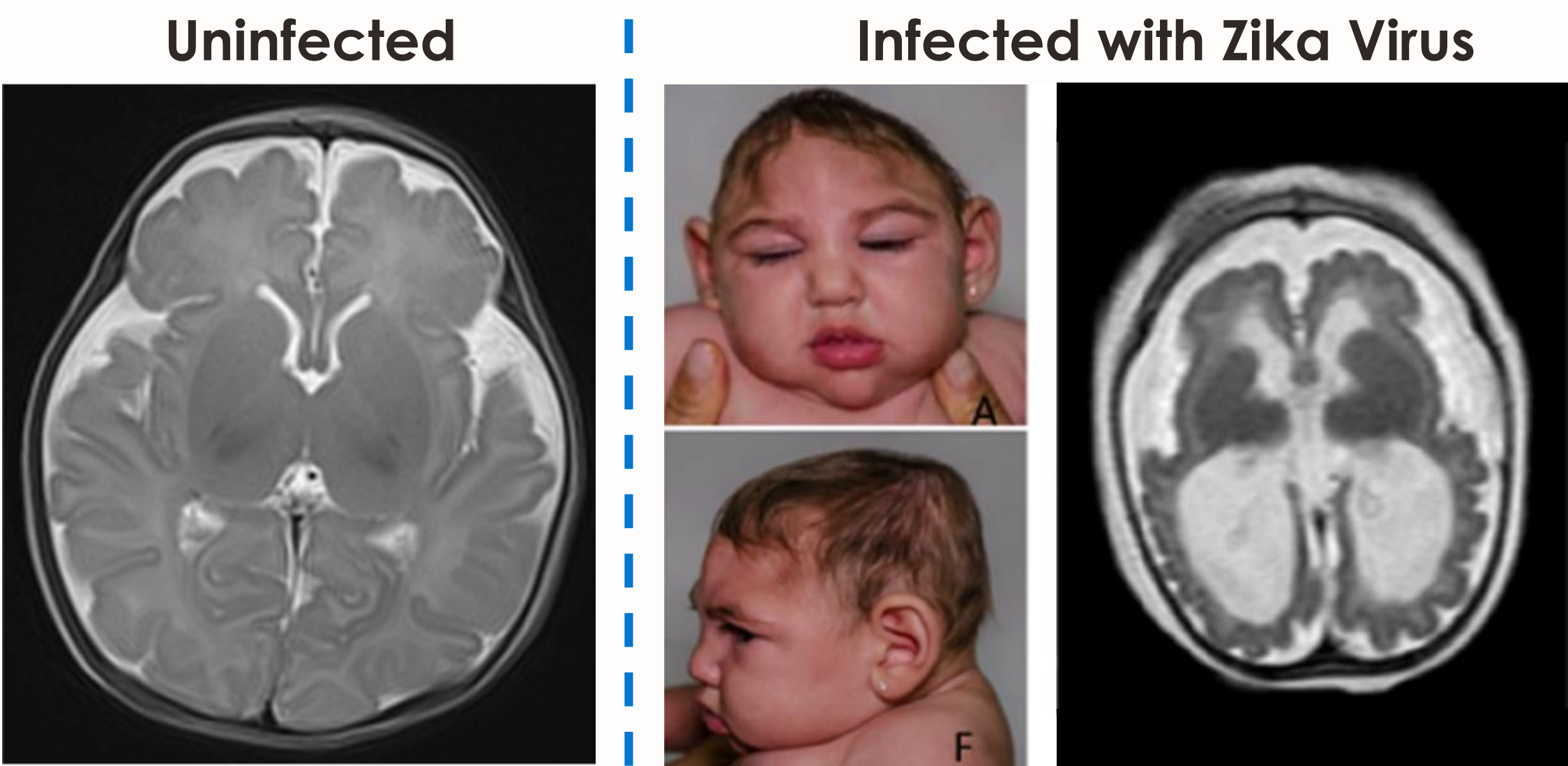


Figure 1. Virally induced brain injury. Prenatal viral infection can cause severe brain injury, as seen by visual examination (collapsed skull) and MRI (tissue loss, images not to scale).³

Objective

Our goal is to identify effective and safe treatments for prenatal Zika infection by repurposing targeted medicines.

Therapeutic Approach

Whole exome genetic sequencing (WES) of severely affected patients identified mTOR (regulator of autophagy) and HEXB (a lysosomal enzyme) as modifiers of injury.⁴ Here, we repurpose and test four drugs that manipulate the autophagy-lysosomal pathway in unique ways; chloroquine, metformin, trehalose, and PD146176. Chloroquine serves as our positive control.⁵

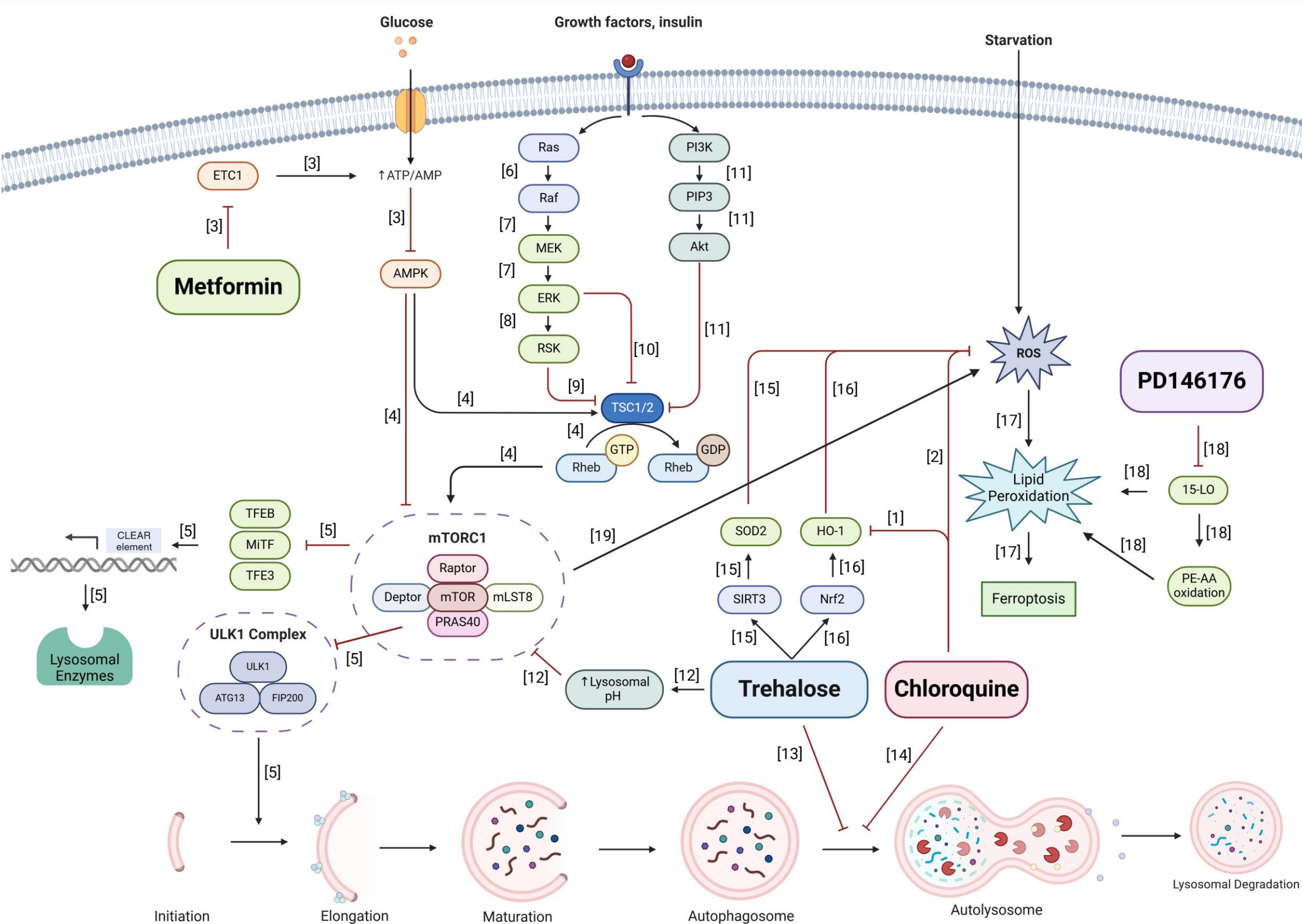


Figure 2. Therapeutic approach. Chloroquine, metformin, trehalose, and PD146176 manipulate autophagy and/or the lysosome by distinct and relevant mechanisms.

Experimental Approach

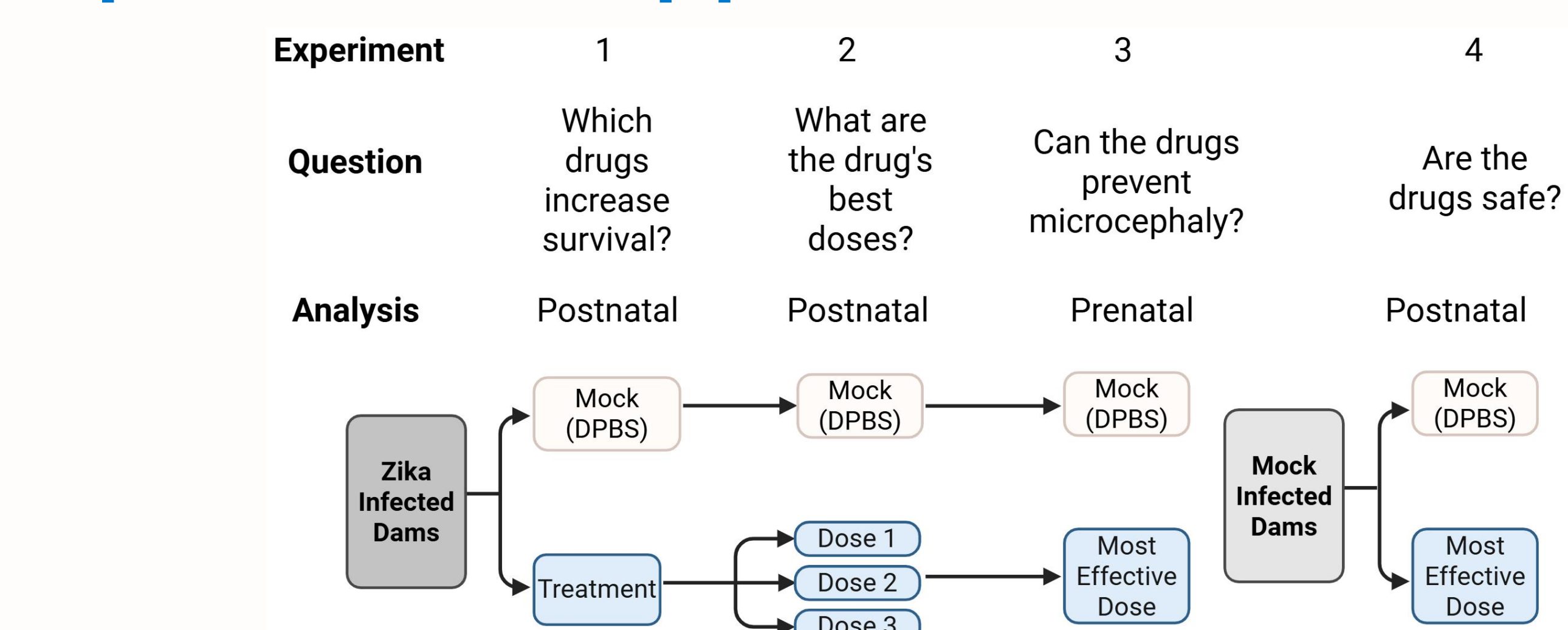


Figure 3. Experimental approach. We outlined four experiments that use mock- and Zika-infected mice to determine the optimal doses of treatments that are effective and safe, as evaluated both prenatally and postnatally.

Methods

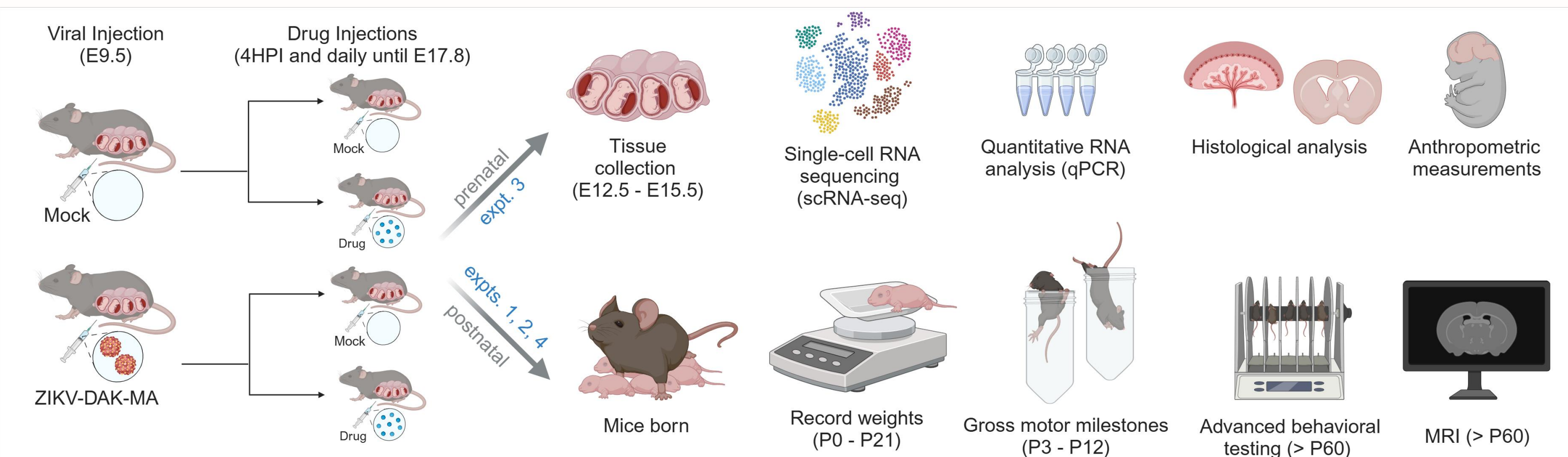


Figure 4. Experimental system. Experimental design of prenatal Zika infection in a humanized immunocompetent (hSTAT2^{KJ/KJ}) mouse model with pipelines for both prenatal and postnatal analyses.

Preliminary Data: Prenatal Viral Infection

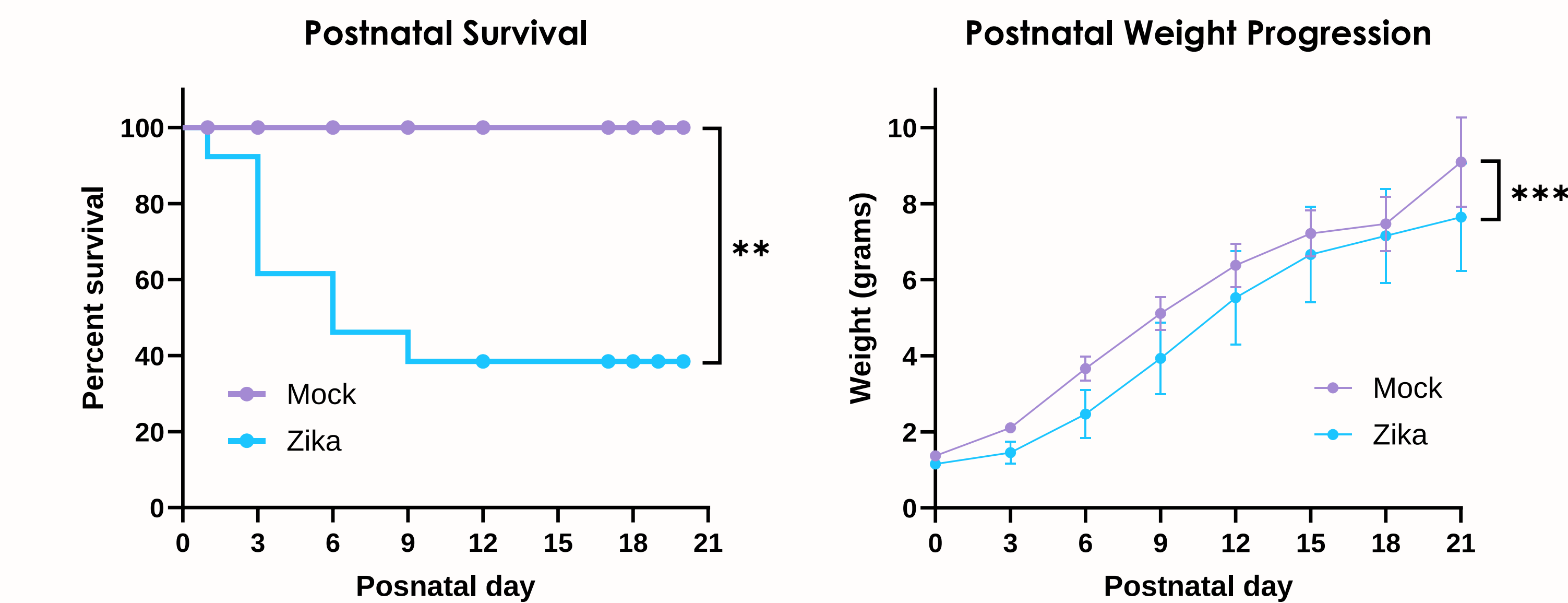


Figure 5. Postnatal survival. Prenatal Zika infection caused 62% death for mice by P21.⁶

Figure 6. Postnatal weight progression. Prenatal Zika infection caused delays in weight gain in mice.⁶

Preliminary Results: Treatments

		Treatment				
		Mock	Chloroquine 50 mg/kg/day	Metformin 100 mg/kg/day	Trehalose 3,333 mg/kg/day	PD146176 80 mg/kg/day
Infection	Mock	n = 1	100, n = 2	-	-	-
	Zika	67, n = 3	0, n = 2	100, n = 1	0, n = 1	-

Table 1. Percentage of successful pregnancies. Prenatal Zika infection can cause spontaneous abortions. This is exacerbated with chloroquine (50 mg/kg/day) and potentially trehalose (3,333 mg/kg/day).

Metformin Increases Survival

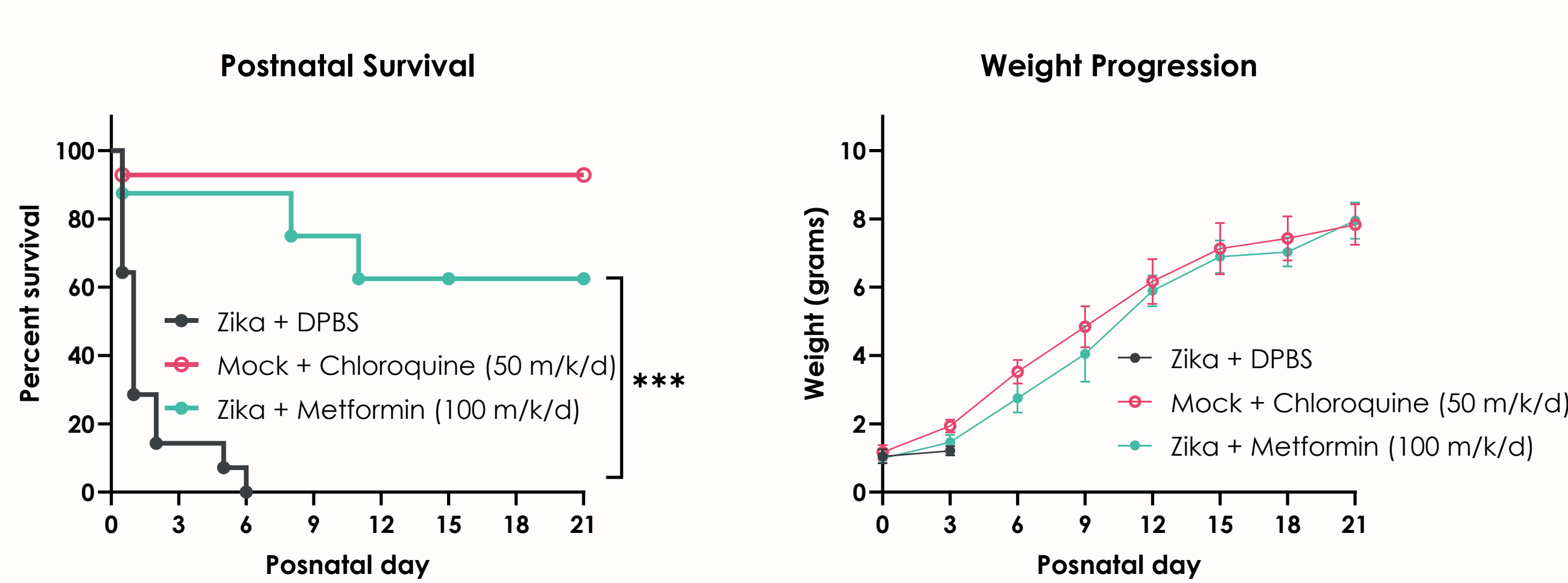


Figure 7. Postnatal survival after treatment. Metformin rescues survival after infection from 0% to 62.5% by P21. Chloroquine causes 93% survival (active data collection).

Figure 8. Postnatal weight progression after treatment. Metformin may rescue weight progression after viral infection. Mock + Chloroquine in active data collection.

Conclusions

Metformin may improve postnatal outcomes for prenatal viral infection. The interaction between chloroquine at 50 mg/kg/day and this viral strain (DAK-MA) may be toxic. Trehalose at 3,333 mg/kg/day may also be toxic. Daily IP injections during pregnancy might cause adverse postnatal outcomes for mice.

Future Directions & Research Timeline

Immediate next steps include growing our sample sizes for each experimental condition, determining the effects of daily IP injections, if any, on outcome, and establishing a dose-dependent curve for each drug. Long-term research plan outlined below.

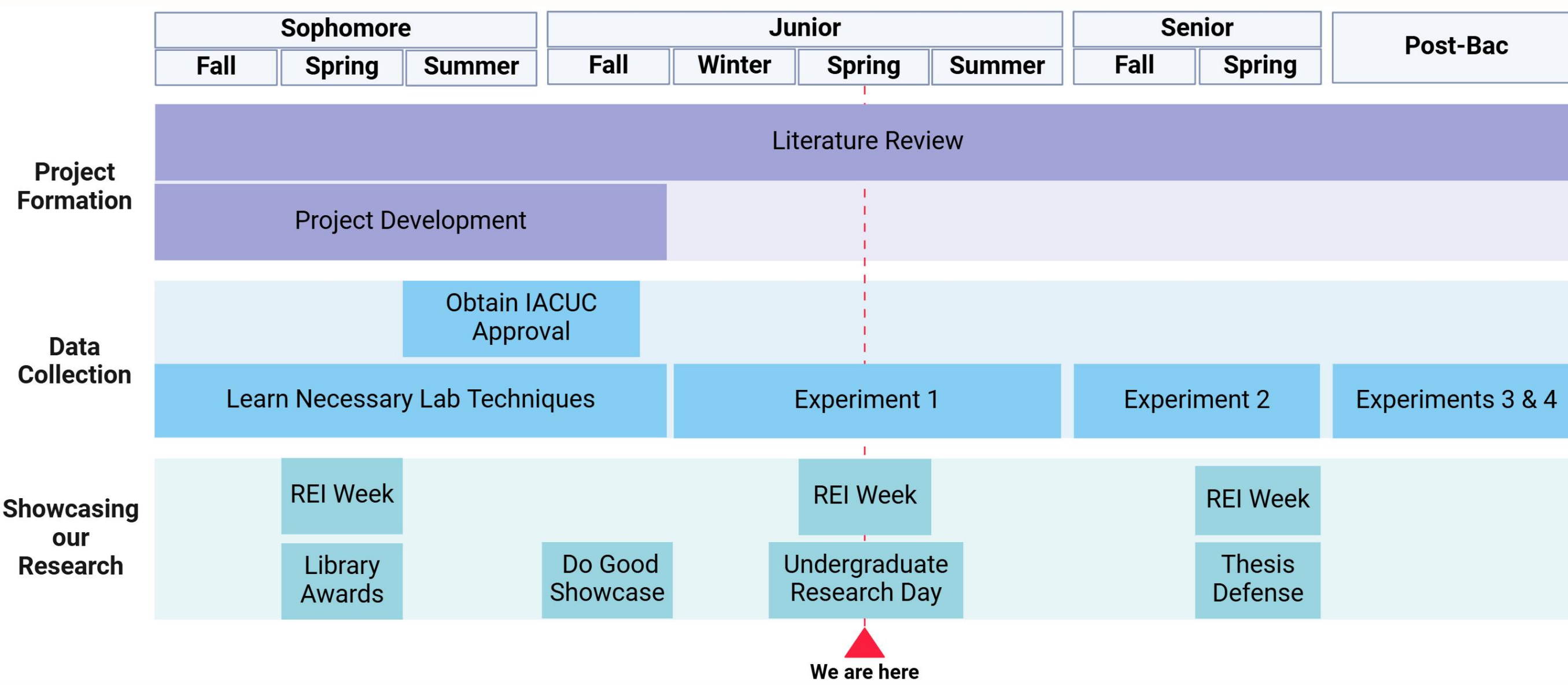


Figure 9. Timeline. We will continue to work on experiments while sharing our work.

Acknowledgments: We would like to thank Dr. Shao, Dr. Zierdan, Dr. Zhang, Dr. Lovell, Dr. Lansverk, and Ms. Tchangalova for their continuous mentorship and support. We would also like to thank the Research Animal Facility staff at Children's National Hospital for their support.

Awards/Funding

- K08NS119882 – NIH
- Intramural Funding – Children’s National Research Institute
- LaunchUMD, University Libraries, Do Good Institute – UMD

Figures: Created with BioRender & GraphPad Prism

References

