

THE NUCLEOLUS IN TULIPA

BY

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THESIS SUBMITTED TO THE FACULTY OF THE GRADUATE SCHOOL
OF THE UNIVERSITY OF MARYLAND IN PARTIAL FULFILL-
MENT OF THE REQUIREMENTS FOR THE DEGREE OF
DOCTOR OF PHILOSOPHY
1936

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THE NUCLEOLUS IN TULIPA

Introduction

Nucleoli apparently occur in the nuclei of practically all living organisms (Sharp (21), Wilson (31)). The possible exceptions are the Myxophyceae and Schizomycetes. Even these may at least possess true nuclei (Smith (24), Tanner (26)) although it is still a question. The universal occurrence of nucleoli has resulted in an extensive examination of this structure in many diverse types of living organisms. In spite of many views that have resulted, the true function and significance of the nucleolus in the economy of the cell is today a matter of opinion. Much has been learned, however, as to the morphological characteristics of this body and its relationships with other nuclear constituents, particularly the chromosomes.

No attempt will be made in this paper to review all the literature on the nucleolus. Montgomery (14) thoroughly reviewed it up to 1897, Wager (30) reviewed a number of the important papers from 1897, and earlier, to 1904. Recent summaries have been presented by Yamaha and Sinoto (32), and Guilliermond and Mangelot (4). Sharp (21) has summarized many of the views regarding the nucleolus and particularly the relation of the chromosomes to this structure.

The significance of the satellited chromosomes in this connection has been a matter of debate since the work of S. Nevaschin (17) who first discovered their existence and relation to the nucleolus in Galtonia Candicans. DeMol (13), Heitz (5,6) McClintock (11,12), and many others, have presented evidence that certain chromosomes have a specific nucleolus-forming function. The last two have shown quite convincingly that it is the chromosomes bearing satellites or certain other similar differentiations that possess this peculiar capacity. Dermen (1), however, seems to be somewhat skeptical of the constancy and importance of the relationship between specific chromosomes and the nucleolus. In evaluating the work in this field it is important to note, as Zirkle (33) has pointed out from the literature and, clearly demonstrated by his own work, that differences in micro-technical methods can lead to radically different fixation images for the nucleolus. Many differences of interpretation have undoubtedly arisen in this way.

The study of nucleoli in Tulipa was undertaken in connection with a morphological study of the chromosomes in this genus. Because of the variable nucleolar numbers, size of the chromosomes, and the occurrence of auto-polyploids, unusually interesting material for an investigation of this kind was available. Nucleolar number, size and, the relation of certain chromosomes to the nucleolus have been considered at different stages of the life cycle.

Materials and Methods

Bulbs were obtained from growers in Europe or in this country. All of the material used had been flowered so its identity was checked. Root-tips were collected in the fall from bulbs grown in the experimental garden. Dual fixations with Flemming's medium solution or Allen and Wilson's Modification of Bouin's fluid were usually made. The ordinary paraffin technic was followed, and iron-alum haematoxylin was used exclusively as a stain for the root-tips. Pre-meiotic and meiotic stages were collected from bulbs kept in a cool storage cellar. The anthers were prepared by the paraffin method or smeared according to the general procedure of Taylor (28) except that ordinary Flemming's medium, and Navaschins fluids were used as fixatives. The crystal violet-iodine-picric acid schedule of Smith (22), or iron-alum haematoxylin were used to stain the smears. Paraffin sections were generally stained in the latter although Flemming's triple stain was occasionally used. Meiotic material was necessarily limited because of the small amount (one flower bud) obtainable from each bulb.

Observations were made with a 2 mm. 90X apochromat oil immersion objective of n.a. 1.30. This was used in conjunction with a 15X compensating ocular. All drawings and

measurements were made with the aid of a camera lucida. Volumetric determinations were recorded in arbitrary scale units (one unit = ap.o.1587 microns). Measurements of nuclei and cells have been recorded in cross-sectional areas in square inches X 2000. A standard planimeter was generally employed in determining these areas, although they were sometimes based on diameter measurements. Photomicrographs were made with the combination described above. Approximate actual magnifications are given for each figure.

Results

Nucleoli in root-tip mitosis

Certain events in the anaphase and, even metaphase, probably influence the development of number of nucleoli in daughter nuclei. The nucleolar matter or plastin (Zirkle (13)), however, appears first in the telophase, and usually disappears in the metaphase.

Anaphase. During late anaphase the chromosomes become inter- and intra-connected by more or less chromatic strands (figs. 1, A and 2, D). These strands, which appear to arise from the matrix, may extend between the proximal and distal arms of the same chromosome as well as between different chromosomes.

Telophase. As the telophase proceeds the chromo-

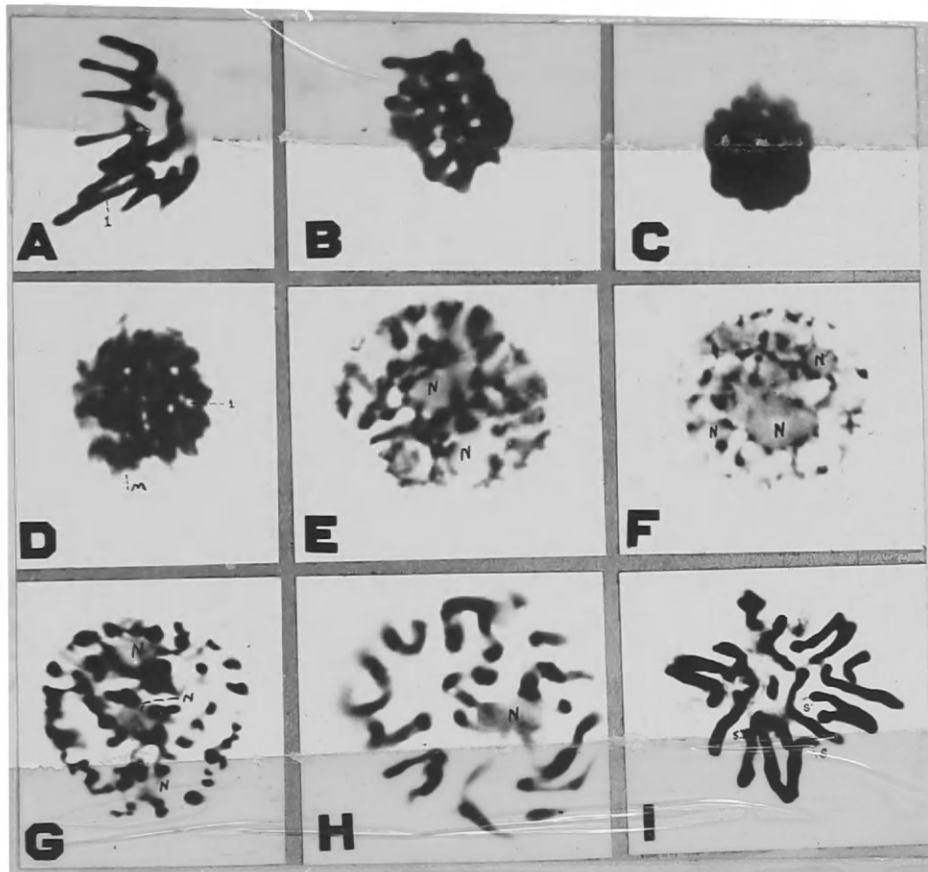


Figure 1. - Development of nucleoli in *T. sprengeri* during mitosis in root-tip: A, Anaphase; B,C,D,E, telophase; F, interphase; G,H, prophase; I, metaphase. In all figs. N denotes plastin and S denotes satellites. X ap. 1800.

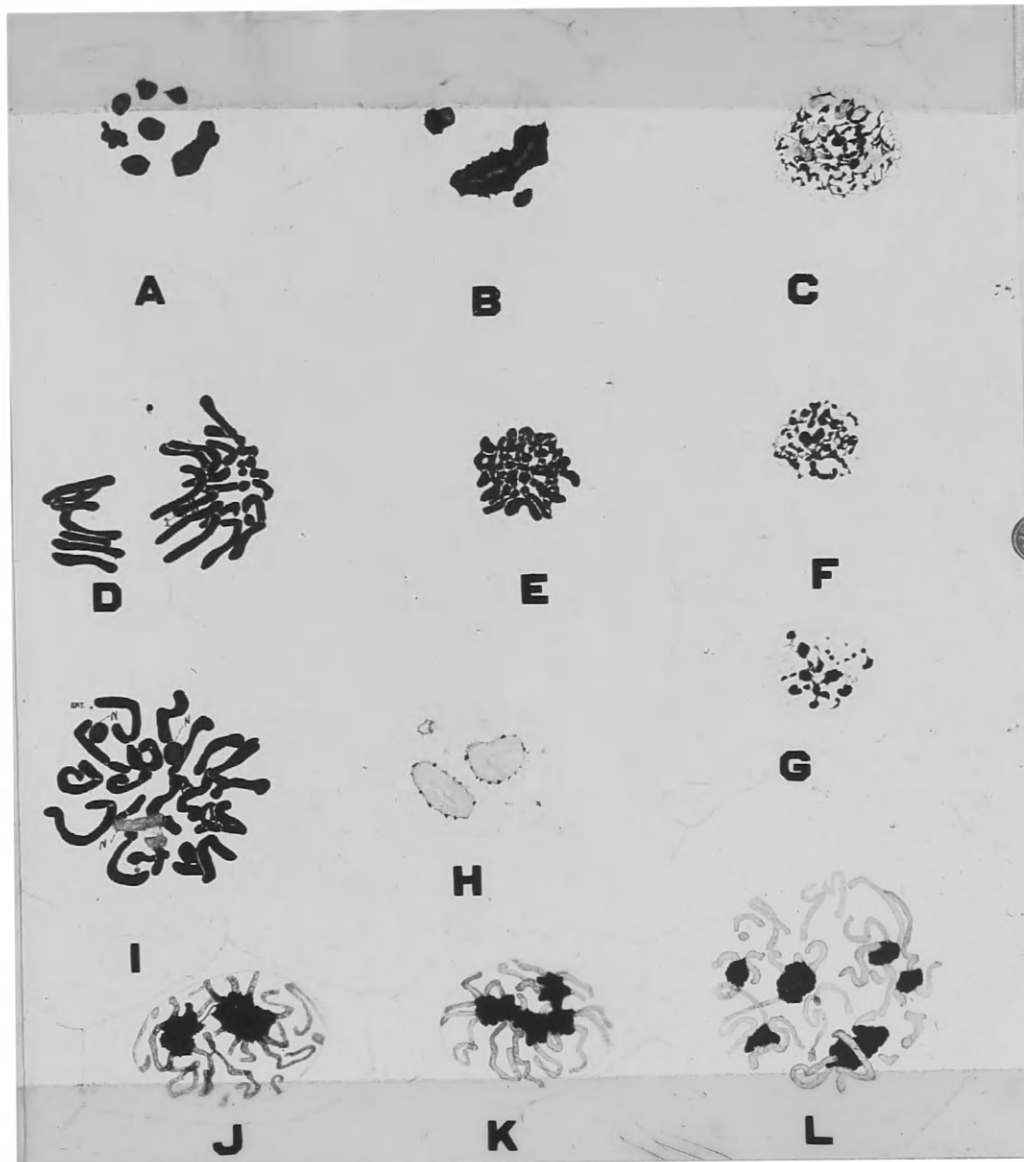


Figure 2. - Development of nucleoli during mitosis in root-tips: A to I, *T. eichleri*: A and B, interphase, Flemming's fixation; C, late telophase, Bouin's fixation; D, anaphase; E, early telophase; F, late telophase, Flemming's fixation; G, telophase later than F, Flemming's fixation; H, interphase, Bouin's fixation; I, early metaphase showing nucleolar remnants (N), Bouin's fixation; J and K, late prophase, *T. linifolia*, Flemming's fixation; L, Carrara, late prophase, Flemming's fixation. All camera lucida drawings X 2000. Reduced ap. $\frac{1}{4}$.

some connecting strands increase in number, and the chromosomes become very closely associated (figs. 1,B, 2,E and 1, C). The first distinguishable plastin appears shortly after the appearance of a membrane-like structure about the mass of chromosomes. Bouin's fixation gives the best telophase differentiation between chromatin and plastin. The latter remains essentially unstained. Early telophase stages are shown in figures 1,E, 2,C and 2 F. Careful studies of many nuclei in different tulips has shown that the appearance of plastin is always associated with the disappearance of chromatin. In many preparations direct connections between the matrix material and developing masses of plastin are indicated. The latter appears to originate at a number of points from the chromosome mass. These areas of collecting plastin may be more numerous than the maximum nucleolar number for the clon (see, nucleolar numbers in root tips, this paper). The developing masses of plastin coalesce to a varying extent as they increase in size. The occurrence of a fixed maximum nucleolar number of each clon examined strongly indicates that some actual flow of the plastin itself must occur in the telophase. Preparations of telophase nuclei frequently give this appearance. During early telophase stages the nucleus is typically irregular in peripheral outline. Toward the end of the telophase the chromatin mass becomes very greatly reduced and the plastin mass

shows further increase. The latter material becomes organized into definite nucleoli. Throughout the progression of the telophase the whole nucleus increases in size, the periphery becoming more regular in outline.

Interphase. During the interphase the chromaticity of the chromonemata remains very low. The latter appear to be arranged radiately about the nucleoli but it is not possible to determine this definitely. The chromonemata are greatly extended and form an interconnected plexus of threads. Irregular small masses of chromatin are usually associated with the chromonematic reticulum (figs. 1, F and 2, H). After fixation in the Bouin's fluid it can be seen that aggregations of chromatin frequently occur on the surfaces of the nucleoli (fig. 2, A and B). The latter frequently show unmistakable signs of their compoundness (as the result of fusion of adjacent nucleoli).

Prophase. As the prophase proceeds the nuclei swell, the chromonemata shorten and, gradually become chromatic. In some tulips, T. sprengeri for example, the chromonemata are very distinctly coiled in the early prophase (fig. 1, G). It soon becomes evident that the developing chromosomes are arranged in bouquet-like fashion about the nucleoli. In the earlier stages of the prophase the doubleness of each chromosome can often be seen. Distinct pairs of chromomeres are also evident. In fig. 20, plate 1,

pairs of chromomeres can be seen in two of the chromosomes, but at one or two points it appears as though certain of the former have not divided. The distinctly radiate arrangement of the chromosomes about the nucleoli is a very characteristic feature of the prophase (figs. 1, H and 2, J, K and L). All of the chromosomes are not always in direct contact with nucleoli. In the nucleus shown in figure 2, J one chromosome was not in contact with a nucleolus but, it was connected to a chromosome that was, by a distinct chromatic strand. This figure shows several other apparently free chromosomes but these were attached at levels not recorded in the drawing. Even though all of the chromosomes do not always touch the nucleoli directly they apparently form a completely interconnected system by virtue of the connecting strands. This seems to be the case in all nuclei examined.

The relations of the chromosome arms to the nucleoli are best studied in preparations fixed in the Bouin's fixative. In figure 22, plate 1, the arrangement of the chromosomes about a prophase nucleolus is shown. Some of the chromosomes are attached to the nucleolus by both of their ends. The small mass of plastin at N, is connected to the distal end of one chromosome and the middle of another chromosome which is also attached to the large nucleolus. It is at similar stages of the prophase that the satellites can sometimes be seen although these structures

are usually hard to demonstrate. Figure 20, plate 1, shows a pair of satellites lying on the surface of a nucleolus although the connecting threads to the end of the chromosome arm could not be distinguished. During the early and middle prophase the satellites appear to be located on the surface of the nucleoli. The chromosomes, whether satellite-bearing or not, also come into contact with the surface or near-surface regions of the nucleolus. In very late prophase or early metaphase the chromosomes may become free of the nucleolus. This seems to be associated with a breaking down or disappearance of the nucleolar matter. In most cases the nucleoli do not persist into metaphase. Sometimes, however, masses of nucleolar matter adhere to the arms of the chromosomes even into metaphase (figs. 15, 16 and 19, pl. 1). The nucleoli appear to remain undiminished in size and staining density up to very late prophase. The final breakdown or disappearance of most of the nucleoli between prophase and metaphase apparently is accomplished rapidly. It does not occur until the chromosome matrices appear to be fully or almost fully developed.

Metaphase. During the metaphase the chromosomes undergo further contraction finally becoming arranged into a definite equatorial plate formation. The satellites are most easily seen during the metaphase but may sometimes be clearly seen in anaphase. In all of the *Leiostemones*

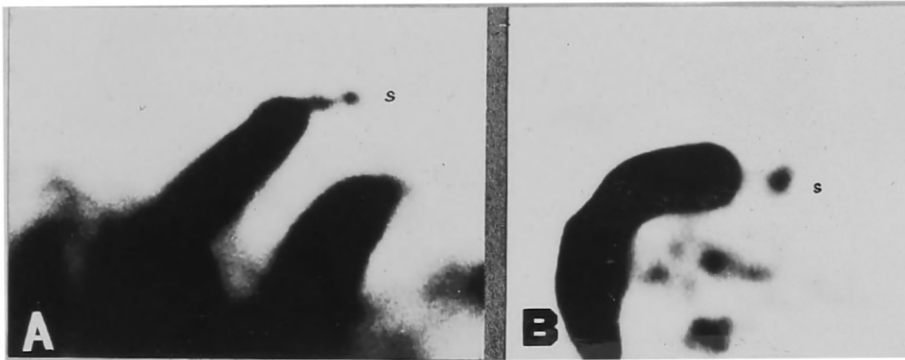


Figure 3. - Metaphase satellites, Darwin: Allard Pierson; A, tandem satellites borne by one J-shaped chromosome; B, normal type of satellite in divided condition at late metaphase. (only distal ends of chromosomes show). Photomicrographs X ap. 6000.

reported in this paper, except T. sprengeri, they occur on the ends of the long or distal arms of the chromosomes. In T. sprengeri (fig. 1, J) the satellites are unusually distinct and are borne only on the short or proximal arms of the chromosomes. In species of the Eriostemones both types of attachment occur. In one Darwin tulip, Allard Pierson, one chromosome bears two satellites in tandem (figs. 3, D and 23, 24, pl. 1). These are apparently very similar to tandem satellites reported in Allium by Taylor (27). Satellites may appear divided before or after the visible separation of the daughter chromatids in the metaphase. In certain tulips demonstration of the satellites in root tip chromosomes is very difficult.

Nucleolar Numbers in Root-tips

Because of the difficulty in demonstrating directly the satellited chromosomes in root-tip nuclei another method of attacking the problem was undertaken. If the maximum number of nucleoli in a given clone is equal to the number of nucleolus-forming chromosomes a mathematical study of nucleolar frequencies should be suggestive. These were determined by random selection of cortical nuclei. Fused nucleoli were counted as single units. Thus the nucleus shown in figure 2, K has but one nucleolus, that in figure 2, A, seven. In determining the characteristic root tip

nucleolar number of any given clon certain precautions were found to be necessary. The characteristic number for a clon can be expressed as the average number of nucleoli per nucleus. In some roots, however, the minimum and maximum nucleolar numbers remain fixed, but the average number per nucleus may show considerable variation at different levels in the same root. In some cases the mean number of nucleoli in the apical region of the root tip is less than in areas farther back from the apex. In some roots the rate of increase appears as an approximately straight line when the mean is plotted against the level in the root in microns. In other cases there is no shift in the mean at successive levels and the graphed line is horizontal rather than inclined. In some roots of T. sprengeri the highest mean occurs nearest the apex.

In order to determine accurately the shift in average nucleolar numbers it was found necessary to average a large number of counts at each level in the root. One section of 20 microns thickness was found insufficient for an accurate determination so two to four sections were counted and the mean level in microns computed for the regions examined. The presence of "false nucleoli" (Zirkle (33)) or non-nucleolar chromatic masses in certain regions of some roots occasionally made accurate counting difficult. In such cases fixation with Allen and Wilson's

Levels on graph (fig.4)	Range in microns in root used in making determinations. The meristematic apex equals 0.	Mean level in microns plotted on graph	Mean no. of nucleoli per nucleus.	Number of nuclei observed
I	0 - 54	27	3.270	100
II	162 - 216	189	3.410	200
III	396 - 450	423	3.660	200
IV	504 - 558	536	3.762	400 (1)
V	630 - 684	659	3.995	200
VI	756 - 810	783	4.165	400 (1)
VII	864 - 918	891	4.247	400 (1)
VIII	990 - 1044	1017	4.410	400 (1)
IX	1098 - 1152	1125	4.500	200
X	1332 - 1386	1359	4.700	200

Table 1. Variations in average number of nucleoli per nucleus in one root of S. L. Carrara at ten different levels.

(1) counts made by counting 100 nuclei in each of two pairs of adjacent sections and then making recounts in same sections and summing both sets of data.

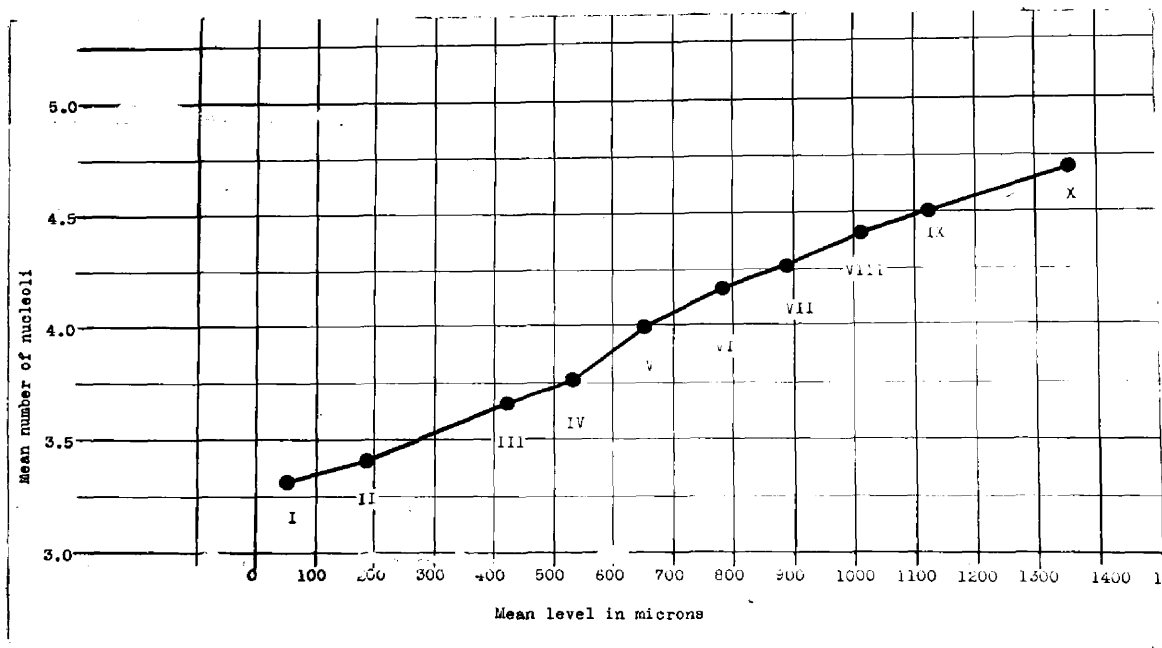


Figure 4. - Relation of level in the root-tip to nucleolar frequency in Carrara. Approximate position of apical histogens equal zero. (See table 1 and text for complete explanation),

modification of Bouin's resulted in a clear differentiation between the true and "false nucleoli". The former were often so lightly stained, however, that the preparations were not used for counting as very small nucleoli might easily have been overlooked. The material used in this investigation was selected with special reference to its desirability for numerical studies. Figure 4 illustrates graphically the gradient in nucleolar mean number obtained at ten different mean levels in a single root of the cottage tulip, Carrara. Table 1 gives the data upon which this graph is based. Extra nucleolar masses of chromatin at level I, made accurate counting difficult. The range of probable greatest accuracy runs from level II to level X. The latter is equal to the "characteristic maximum area".

In figure 5 the nucleolar gradients for several species and varieties are compared. Only a few points have been determined in most cases and the exact slopes are therefore not shown on this graph. The data, however, illustrates the necessity of considering nucleolar number gradients in attempting to determine the characteristic mean nucleolar frequency per nucleus for a clone.

In the tulips examined in this study it was found that if successive nucleolar counts were made in 100 or more nuclei at any one level, or closely adjacent levels, there was often an unmistakable regularity in the frequencies for

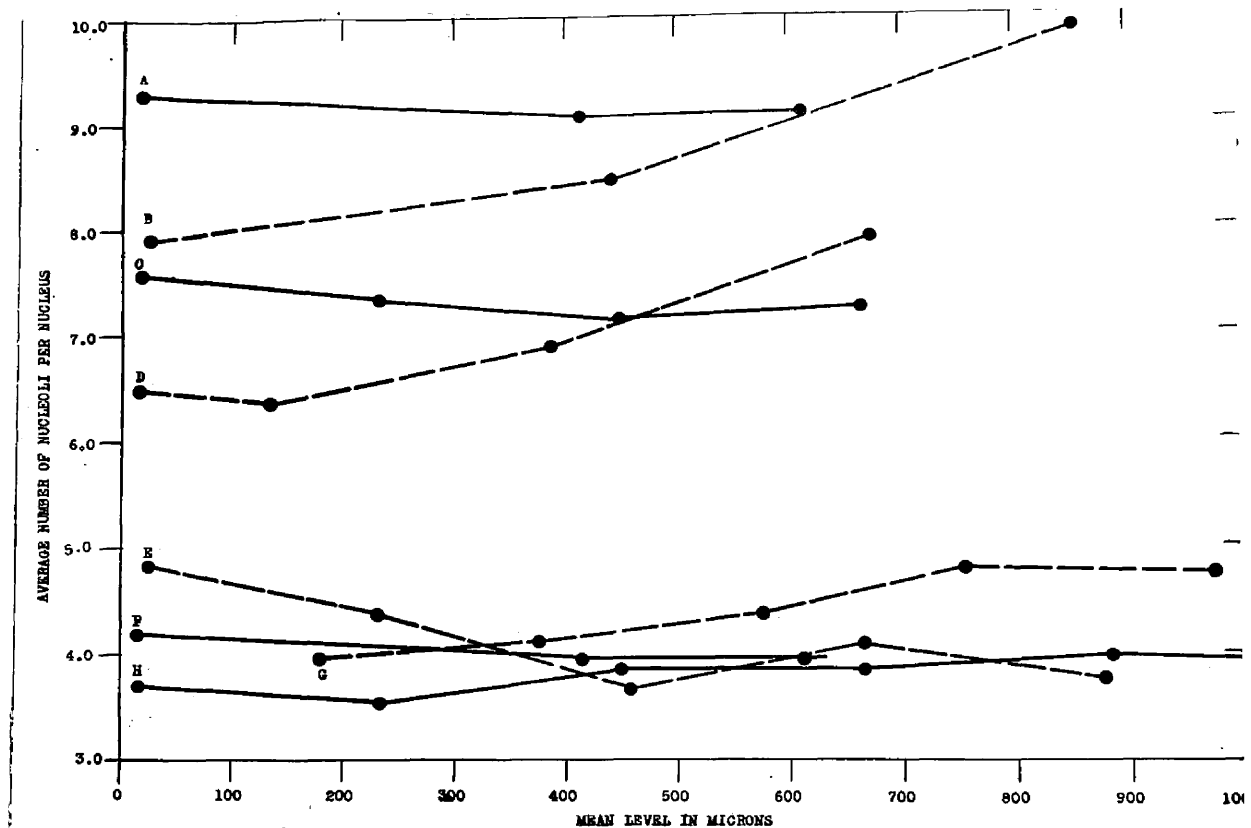


Figure 5. - Relation of level in the root-tip to nucleolar frequency in several species and varieties of *Tulipa*: A, *T. orphanidea* 4n (238 nuclei); B, *T. clusiana* 5n (300 nuclei); C, *T. saxatilis* 3n (273 nuclei); D, Inglescombe Yellow 3n (350 nuclei); E, *T. sprengeri* 2n (580 nuclei); F, *T. hageri* 2n (228 nuclei); G, Mrs. Moon 2n (1000 nuclei); H, *T. fosteriana* 2n (709 nuclei). One root-tip was used in each case.

the different numbers of nucleoli per nucleus. From the distributions obtained it is certain that the nucleolar numbers of at least some of the investigated clons are normally fixed within definite limits in the root tips. This is probably the case in all of the tulips investigated in this study. Even though the mean number of nucleoli per nucleus may shift at different levels in the roots of a given clon the maximum and minimum numbers remain fixed. This is best evidenced by the shapes of the curves obtained when the number of nucleoli are plotted against the percentage of nuclei obtained for each number class. Figures 6 and 7 show the curves obtained for the two diploid cottage tulips Mrs. Moon and John Ruskin. In each figure curves have been plotted for two different levels in the root. Figure 7 also shows the curve obtained by combining the nucleolar counts in 1000 nuclei from two other roots of John Ruskin. Figure 8 shows the curves obtained by plotting the nucleolar frequencies at four of the levels in the root of Carrara used in plotting figure 4. The nucleolar frequency curve for 1665 nuclei, included in both minimal and maximal nucleolar levels of five roots of T. sprengeri, is shown in figure 9.

In each of the above mentioned four tulips the somatic chromosome number is 24. The maximum root-tip nucleolar number in each is obviously eight. In John Ruskin

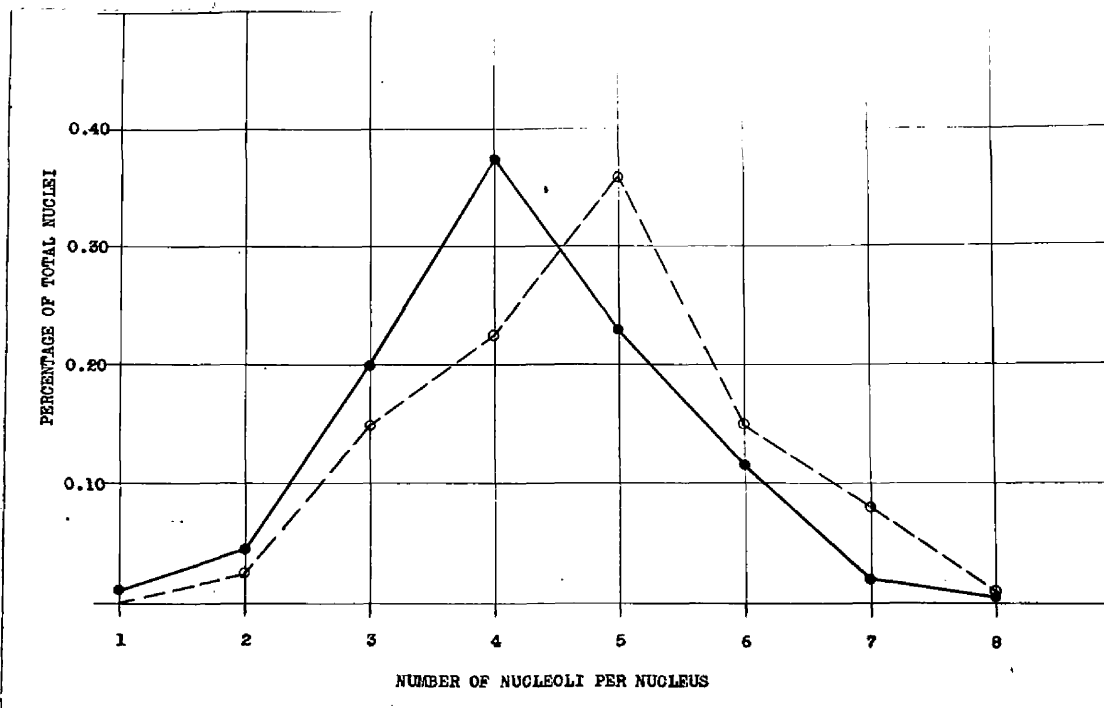


Figure 6. - Nucleolar frequencies at two different levels in a root-tip of Mrs. Moon. Solid line, mean level of 378 microns; broken line, mean level of 972 microns. Each distribution is based on counts of 200 nuclei.

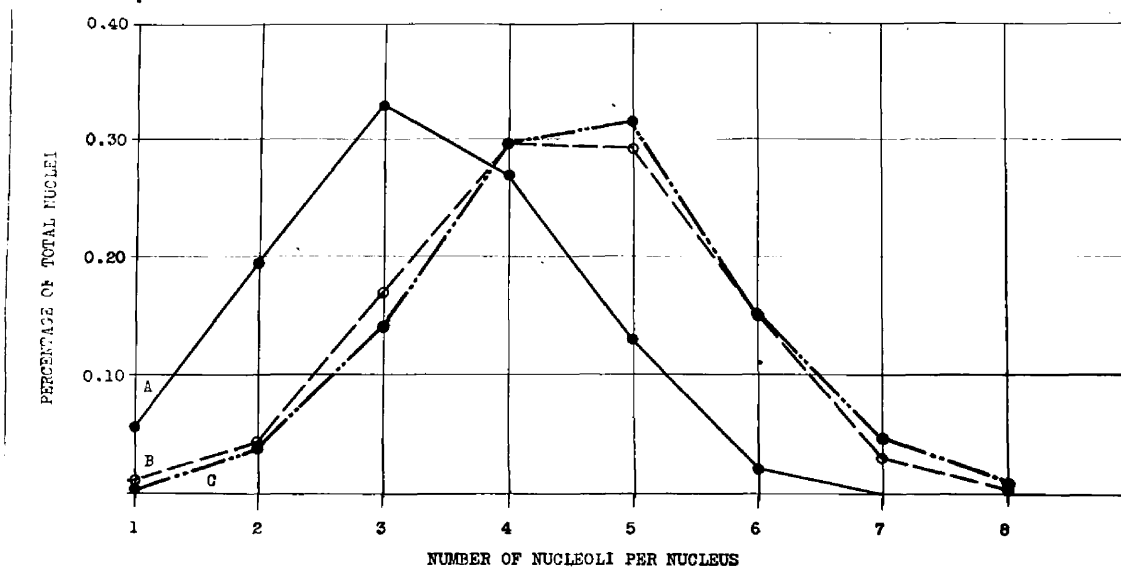


Figure 7. - Nucleolar frequencies in root-tips of John Ruskin: A, root 1, mean level of approximately 380 microns, 200 nuclei; B, root 1, mean level of approximately 980 microns, 300 nuclei; C, based on 1000 nuclei from two roots, counts made at or near levels having maximum characteristic nucleolar averages.

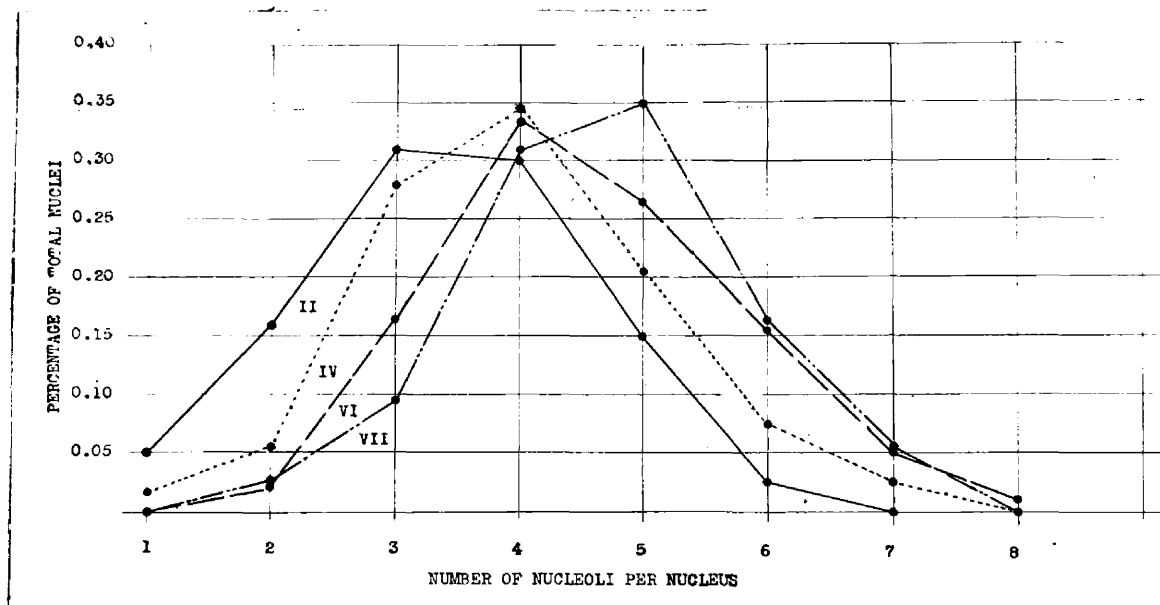


Figure 8. - Nucleolar frequencies at four different levels in a root-tip of Carrara. Roman numerals refer to levels and correspond to table 1.

nucleolar counts have been made in a total of 4585 nuclei, in Carrara at least 4000 and, in Mrs. Moon 2650. The maximum nucleolar number given above, was never exceeded. In addition to the counts actually recorded many others have been made in connection with other studies in the same tulips. No exceptions to the rule just given were found. Other tulips were found to be characterized by maximum nucleolar numbers greater or less than eight. Figure 10 shows nucleolar distribution curves for the triploid tulips Inglescombe yellow, Inglescombe pink and T. lanata. Note that the rates of slope indicate a nucleolar maximum near 13.

It was found by experiment, that merely by inspection of several levels in a root the areas having the highest mean numbers per nucleus can be determined with considerable accuracy. If counts are made in such areas in different roots a characteristic average nucleolar number for any given clon of a species can be determined. Each of the named "garden varieties" represents a clon derived from a single seed or bulb so a characteristic nucleolar number can be calculated for each one of these. While there is some variation, enough counts at comparable levels in different roots give surprisingly uniform values. Table 2 lists the observed nucleolar numerical characteristics for about 30 species, varieties and aneuploid hybrids of Tulipa. One heteroploid

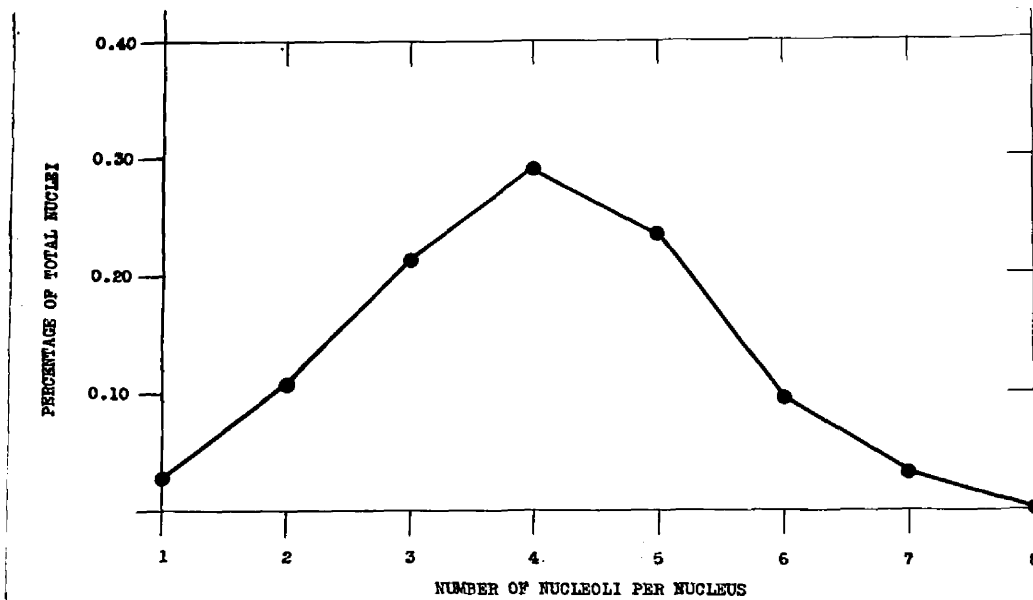


Figure 9. - Nucleolar frequencies in four root-tips of T. sprengeri. Based on 1665 nuclei from different levels.

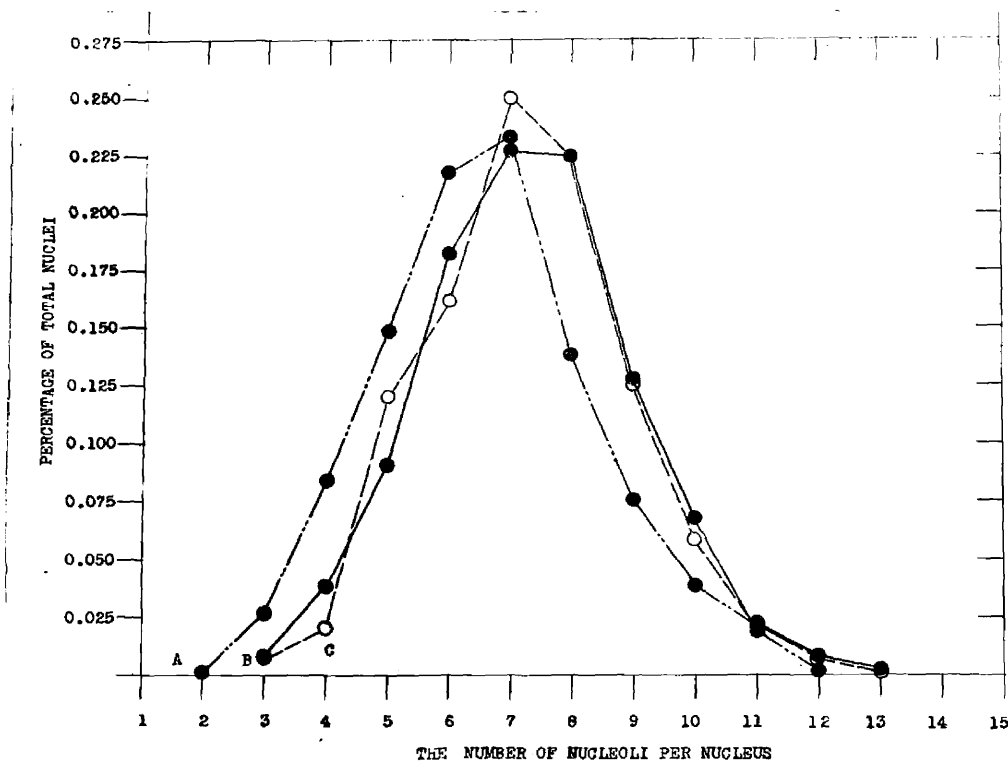


Figure 10. - Nucleolar frequencies in root-tips of three triploid tulips: A, Inglescombe Pink, 488 nuclei from four roots; B, Inglescombe Yellow, 1000 nuclei from two roots; C, T. lanata, 391 nuclei from four roots. (All counts made at levels in the roots having maximum characteristic nucleolar averages for the clons concerned.)

Name and Chromosome number	Avg. number nucleoli each root (roots 1 to 6)						Avg. all roots	Ex. no.	No. nuclei observ- ed
	No.1	No.2	No.3	No.4	No.5	No.6			
<i>T. linifolia</i> Regel (24)	4.07	3.88	4.01	3.76	--	--	3.83	3	442
<i>T. stellata</i> Hook (24)	4.77	4.47	4.50	4.92	4.52	4.51	4.52	3	204
<i>T. fosteriana</i> Hook (24)	4.91	4.35	4.41	4.53	--	--	4.52	1	409
<i>T. sorengeri</i> Baker (24)	3.72	3.78	4.00	---	--	--	3.73	1	1222
<i>T. hageri</i> Heldr. (24)	4.00	3.91	--	--	--	--	3.95	3	531
<i>T. violacea</i> Boiss. (24)	3.84	3.36	--	--	--	--	3.7	7	352
<i>S. L. Carrara</i> (24)	4.56	4.51	4.65	4.65	4.70	--	4.50	3	542
<i>S. L. Mrs. Moon</i> (24)	4.74	4.71	4.73	4.73	--	--	4.73	3	540
<i>S. L. John Ruscin</i> (24)	4.84	4.77	4.76	4.76	--	--	4.76	3	415
<i>S. L. Adonis</i> (24)	5.97	5.80	5.80	5.81	5.79	5.79	5.79	3	1700
New Mendel; Herman Waller (24)	4.71	4.65	4.65	4.65	--	--	4.65	7	1941
Breeder; Louis XIV (24)	5.34	5.35	5.35	5.35	--	--	5.37	9	472
New Mendel; Gipsy (24)	4.60	4.70	4.70	4.70	--	--	4.65	2	112
S.E. La Reine Maximus (24)	5.44	--	--	--	--	--	5.44		100
DO 72 chromosome area	22.35	--	--	--	--	--	22.35	1	17
<i>T. lanata</i> (36)	7.13	7.62	7.34	7.67	--	--	7.44	13	211
<i>T. sentillis</i> Sieber (36)	7.35	7.93	7.93	--	--	--	7.40	10	211
<i>T. L. Inlescombe rim</i> (36)	6.32	6.51	6.81	6.77	--	--	6.60	12	380
<i>T. L. Inlescombe Yellow</i> (36)	7.91	7.51	7.52	7.92	--	--	7.70	13	475
<i>T. L. sentilla</i> Boiss. (48)	5.28	5.67	5.27	5.82	--	--	5.35	13	124
<i>T. L. estonica</i> Regel (48)	7.05	--	--	--	--	--	7.05	11	211
<i>T. L. oromoides</i> Boiss. ? (48)	9.14	8.82	--	--	--	--	8.98	15	317
<i>T. alusiana</i> DC. (60)	9.20	9.53	9.54	10.0	--	--	9.57	17	104
Louis XIV x Mrs. Moon (24)	4.40	--	--	--	--	--	4.40	3	101
Louis XIV x Mrs. Moon (24)	4.35	--	--	--	--	--	4.35		100
Carrara x John Ruscin (24)	4.76	--	--	--	--	--	4.76		100
Carrara x John Ruscin (24)	4.43	--	--	--	--	--	4.43		100
Inlescombe Yell. x John Ruscin (27)	4.60	--	--	--	--	--	4.60	11	200
Inles. Yell. x John Ruscin (27)	4.45	--	--	--	--	--	4.45	3	91
Inles. Yell. x John Ruscin (30)	5.32	--	--	--	--	--	5.32	10	17
Inlescombe Y. x John Ruscin (30)	5.45	--	--	--	--	--	5.45	10	17

Table 2. Nucleolar numbers in *Tulipa*.

For graphical representation of some of this data see figures 10b and 10c.

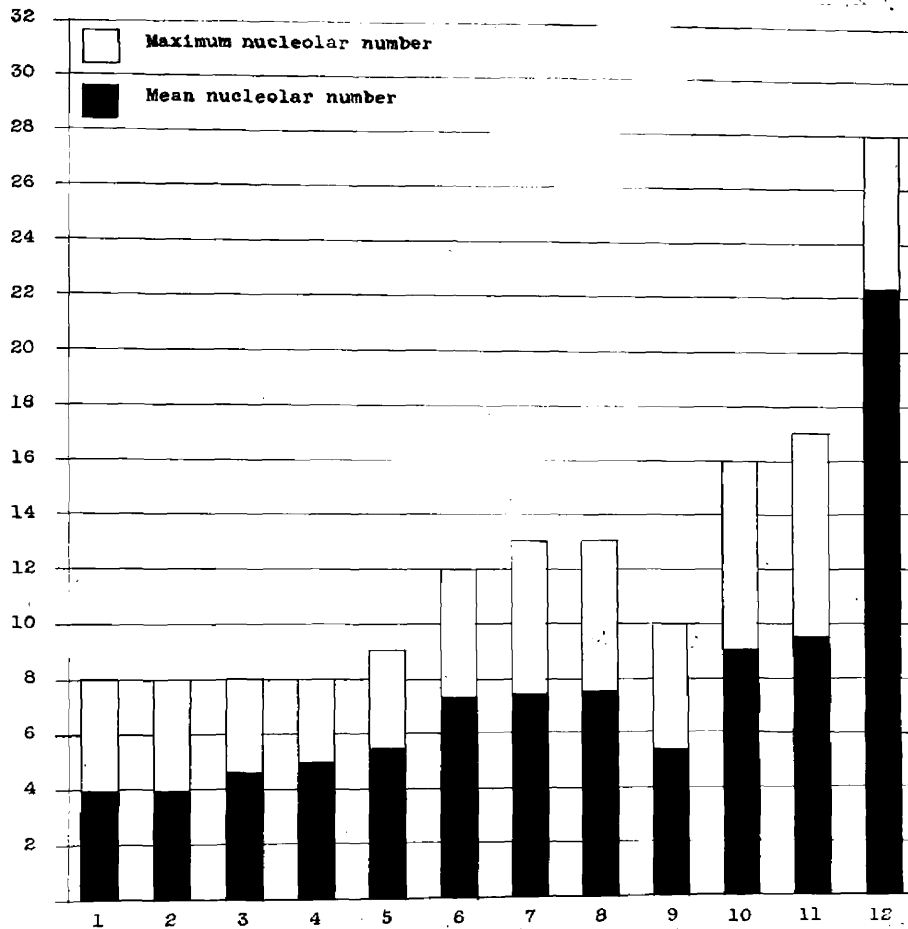


Figure 10b. - Nucleolar numbers in species and varieties of Tulipa:

- | | |
|-------------------------|---------------------------|
| 1-T.linifolia (24) | 7-T.lanata (36) |
| 2-T.hageri (24) | 8-Inglescombe Yellow (36) |
| 3-T.fosteriana (24) | 9-T.chrysantha (48) |
| 4-John Ruskin (24) | 10-T.orphanidea (48) |
| 5-La Reine Maximus (24) | 11-T.clusiana (60) |
| 6-T.saxatilis (36) | 12-La Reine maximus (72 ? |

(somatic chromosome numbers in brackets after names)

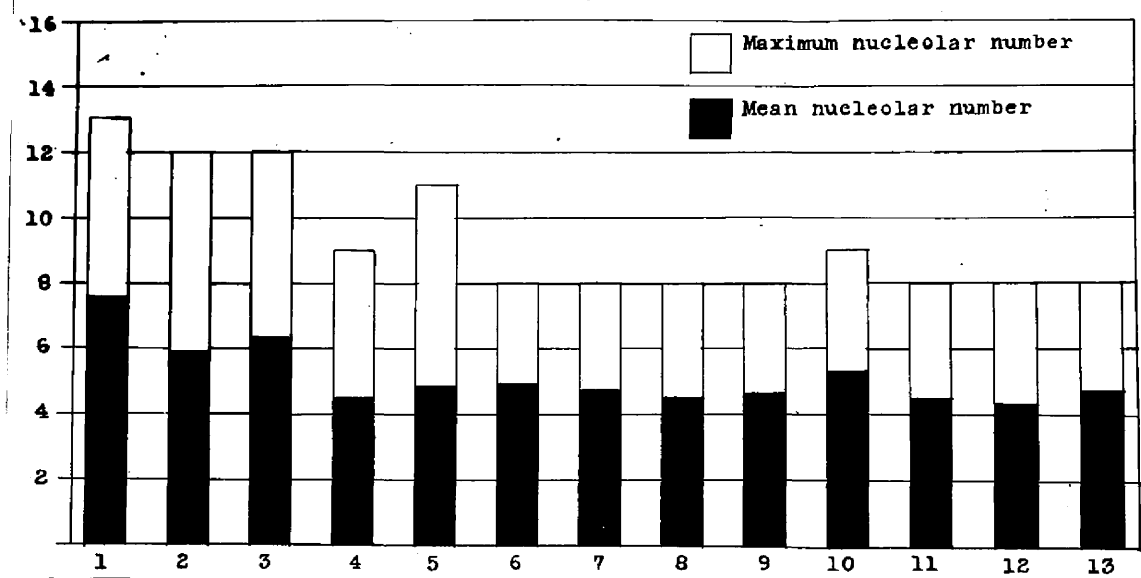


Figure 10c.- Nucleolar numbers in tulip varieties and their hybrids:

- 1 - Inglescombe Yellow (36)
 - 2 - Inglescombe Yellow X John Ruskin, hybrid no.1 (32)
 - 3 - Inglescombe Yellow X John Ruskin, hybrid no.2 (30)
 - 4 - Inglescombe Yellow X John Ruskin, hybrid no.3 (28)
 - 5 - Inglescombe Yellow X John Ruskin, hybrid no.4 (27)
 - 6 - John Ruskin (24)
 - 7 - Carrara X John Ruskin, hybrid no.1 (24)
 - 8 - Carrara X John Ruskin, hybrid no.2 (24)
 - 9 - Carrara (24)
 - 10--Louis XIV (24)
 - 11- Louis XIV X Mrs. Moon, hybrid no.1 (24)
 - 12- Louis XIV X Mrs. Moon, hybrid no.2 (24)
 - 13- Mrs. Moon (24).
- (Somatic chromosome numbers are given in the brackets)

chimera of a diploid root of La Reine Maximus is also included. In this abnormal case one excellent metaphase plate contained 72 chromosomes (fig. 11,). The nucleolar counts were made in cells closely adjacent. The autoheteroploid nuclei were readily distinguished (fig. 11,) by their great size. They probably had the same chromosome number as the one cell in which the count was made but, in any case it must have been considerably more than 24.

Volumetric Relationships in the Root-tip

Measurements were made of cell, nuclear, and nucleolar sizes to determine their relation, if any, to nucleolar number. It was thought the nucleolar number gradients, previously referred to, might be explained in part on this basis. Measurements were confined to the mitotic regions although in the upper limits very few cell divisions occurred.

Nucleolar number in relation to cell and nuclear size. Two methods were employed. First an attempt was made to correlate nuclear and cell size with nucleolar number at a given level in the root, and second to make comparisons of these factors between several different levels in the same root. Roots of Carrara and T. sprengeri were studied in detail because of the radically different types of nucleolar number gradients obtained in each. In Carrara the highest mean number of nucleoli occurred farthest from the apex.

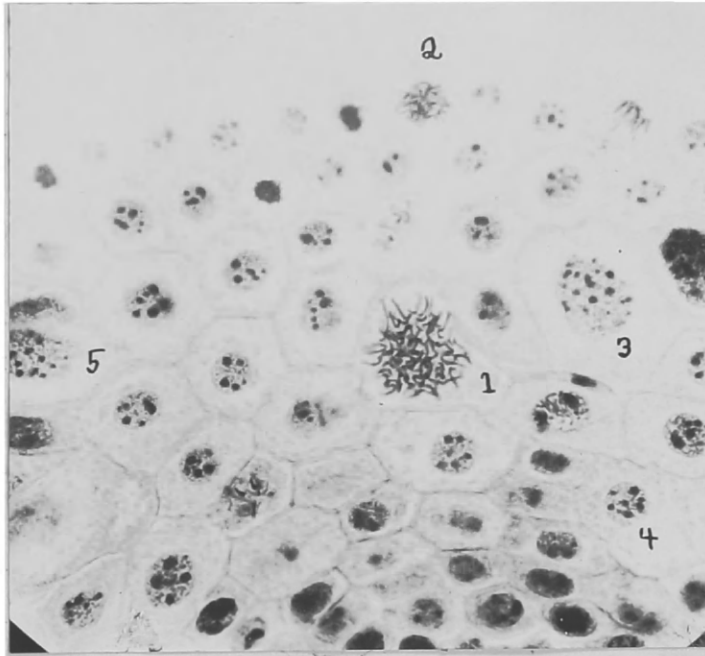


Figure 11. - Sector of a root-tip of La Reine Maximus showing chimeral areas. The metaphase plate at (1) has 72 chromosomes, the plate at (2) has 24 chromosomes. Heteroploid nuclei (with 72? chromosomes) are shown at (3) and (5). A diploid nucleus is shown at (4) and elsewhere. Nucleolar counts were made in nuclei above and below the regions marked (1) and (3). Photomicrograph X ap. 500.

In T. sprengeri the highest nucleolar frequencies occur near the apex, the rest of the mitotic region having a lower, and fairly constant average number of nucleoli. In table 3 the measurements of 432 nuclei in one root of Carrara are recorded with their nucleolar numbers. From this data there seems to be little if any correlation between nuclear size and nucleolar number. If the mean nuclear sizes for all levels are considered, however, there does appear to be some slight increase in nuclear size as the nucleolar number increases (fig. 12). The average size for nuclei containing six nucleoli is approximately the same as that for two nucleolate nuclei, however. This data clearly shows that the nuclei are larger at the 423 micron level than at either the 189 or 891 micron levels. A similar situation seems to hold in the other roots examined i.e., the maximum average nuclear size occurs somewhere between the apex and the upper limits of the mitotic region.

The situation with respect to nuclear size and nucleolar number relationships in T. sprengeri is somewhat different than in Carrara. In the former the smaller nuclei tend to have the higher nucleolar numbers. There is some over-lapping, however, in sizes of individual nuclei between the different nucleolar number classes. The maximum nucleolar number may occur in a large nucleus or a small one. The same is true for the lower nucleolar numbers. That the smaller nuclei generally contain the largest number of nucleoli,

Nucleolar Number	Nuclear size (average)(1) at three different mean levels			Avg. nuclear size all levels	No. of nuclei measur- ed
	189 microns	423 microns	891 microns		
1	0.93	1.18	-----	1.05	15
2	1.03	1.46	0.97	1.15	65
3	1.14	1.46	1.09	1.23	88
4	1.10	1.49	1.13	1.24	95
5	1.24	1.46	1.09	1.26	95
6	1.10	1.34	1.07	1.17	56
7 & 8	-----	1.47	1.07	1.27	18
Aug.	1.09	1.40	1.08	1.19	432

Table 3. Nuclear size in relation to nucleolar number in one root of S.L. Carrara (See table 1.) (1) Nuclear sizes expressed as square inches X 2000.

Cell size(1)	Nuclear size	No. of nucleoli	Total nucleolar volume (2)	No. cells measured	Mean Level in root in microns	Minimum nucleolar volume	Maximum nucleolar volume	Size of cell with minimum nucleolar volume	Size of nucleus with maximum nucleolar volume	Size of cell with maximum nucleolar volume	Size of nucleus with maximum nucleolar volume
4.30	1.63	2	37,577.1	10	423	26,437.6	49,998.6	3.46	1.27	5.31	2.00
4.66	1.70	4	35,941.2	10	423	24,844.5	50,507.1	4.39	1.62	4.19	1.79
4.69	1.60	6	29,086.8	10	423	18,543.0	50,999.7	2.58	1.30	6.43	2.38
6.20	1.07	2	14,140.7	5	891	7,726.3	22,194.0	5.91	1.12	6.98	0.98
7.51	1.26	4	13,972.7	10	891	10,041.6	19,526.6	8.27	1.31	9.20	1.46
7.93	1.14	6	9,967.6	10	891	6,797.2	14,458.2	7.05	1.00	9.70	1.35

Table 4. Nucleolar number and volume in relation to nuclear and cell size at two different levels in one root (see table 1.)

(1) Cell and nuclear sizes expressed as square inches X 2000.

(2) Nucleolar volumes expressed in cubic scale units.

Nucleolar Number	Nuclear sizes (1) at different levels in two roots						Root No. 1		Root No. 2	
	Mean levels in Microns root no. 1				Mean levels in microns root no. 2.		Avg. nuclear size all levels	Number of nuclei measured	Avg. nuclear size all levels	Number of nuclei measured
	27	243	459	891	27	441				
1	-----	1.03	1.05	0.95	0.93	1.09	1.01	21	1.01	18
2	0.83	1.02	1.03	1.07	1.02	1.09	0.98	50	1.05	43
3	0.80	0.99	1.03	0.98	0.97	1.10	0.95	78	1.03	70
4	0.79	0.93	0.99	0.95	0.93	1.03	0.91	65	0.98	57
5	0.72	0.88	0.89	0.86	0.95	0.99	0.83	83	0.97	50
6	0.72	0.91	0.91	0.80	0.89	0.99	0.83	57	0.94	21
7	0.68	0.79	0.83	0.80	0.88	0.97	0.83	37	0.92	15
8	0.67	-----	-----	0.80	0.82	-----	0.73	8	0.82	2
Aug.	0.74	0.93	0.96	0.90	0.92	1.03	0.88	399	0.97	276

Table 5. Nuclear size in relation to nucleolar number at different levels in two roots of T. sprengeri.

(1) Nuclear sizes expressed as square inches X 2000.

Avg. level in the root in microns	No. of nucleoli	Avg. cell size (1)	Avg. nuclear size (1)	Avg. total nucleolar volume (2)	Maximum nucleolar volume	Minimum nucleolar volume	Size of cell with maximum nucleolar volume	Size of nucleus with maximum nucleolar volume	Size of cell with minimum nucleolar volume	Size of nucleus with minimum nucleolar volume	No. of cells measured
27	3	2.21	0.83	9,236.8	12,782.9	4,374.1	2.24	0.97	1.70	0.61	10
DO	7	1.91	0.77	6,098.6	10,733.6	3,352.7	2.54	0.79	1.70	0.60	10
459	1	4.33	1.16	33,609.8	47,840.6	28,807.8	4.20	1.24	3.78 & 5.45	1.11 & 1.22	8
DO	2	4.08	1.18	25,532.2	46,842.2	18,219.5	4.46	1.35	3.41	1.10	13
DO	3	3.43	1.04	16,532.8	22,519.2	8,330.1	4.39	1.13	2.89	0.98	10
DO	4	3.95	1.14	21,960.7	33,690.2	12,301.9	6.11	1.47	2.45	1.07	10
DO	5	3.18	0.93	14,967.6	27,528.4	9,951.3	2.80	1.05	3.02	0.80	10
DO	6	2.77	0.90	12,879.3	21,740.6	7,306.4	3.46	1.22	2.00	0.72	10
1368	1	5.75	0.93	17,407.9	36,835.2	11,777.0	7.49	1.04	8.18	0.84	10
DO	2	5.92	0.90	13,406.0	17,721.2	6,617.3	4.17	0.92	5.30	0.75	10
DO	3	4.92	0.87	11,514.6	20,441.6	4,873.2	5.08	1.07	2.16	0.90	10
DO	4	5.81	0.91	11,401.0	13,998.6	8,828.6	4.70	0.92	4.07	1.00	10
DO	5	5.04	0.82	8,661.1	11,363.5	5,515.6	8.23	0.92	3.00	0.88	11
DO	6	4.31	0.84	8,223.9	13,708.8	4,749.7	6.07	0.99	4.78	0.70	10

Table 6. Nucleolar number and volume in relation to nuclear and cell size at three different levels in one root of T. sprengeri.

(1) Size expressed as square inches X 2000.

(2) Volume in cubic scale units.

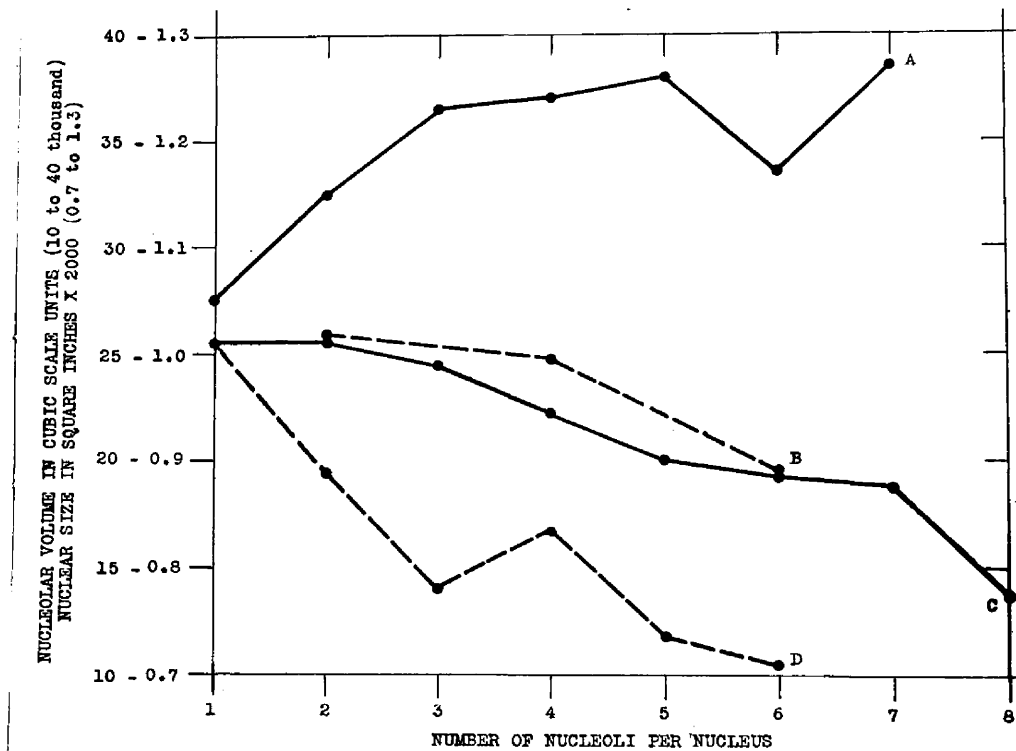


Figure 12. - Relation between nuclear size, nucleolar volume and nucleolar number in root-tips of T. sprengeri and Carrara: A, Nuclear size in Carrara; B, nucleolar volume in Carrara; C, nuclear size in T. sprengeri; D, nucleolar volume in T. sprengeri.

however, is clearly shown by the data recorded in tables 5 and 6. In figure 12 the average nuclear sizes for all levels in both roots of T. sprengeri recorded in table 5 have been plotted against the number of nucleoli. It can be seen that the rate of decrease in nuclear size is not the same between all intervals.

Nucleolar, Nuclear and cell volumes. Measurements made in roots of Carrara and T. sprengeri show that large nuclei generally have larger nucleolar volumes than small nuclei in the same sections. Tables 4 and 6 give the nuclear and cell sizes together with the nucleolar volumes for several levels in one root for each tulip. In each case there is only one exception to the rule that the highest observed nucleolar volumes occur in nuclei larger than those containing the corresponding minimum nucleolar volumes. The large nuclei usually occur in cells of greater cross-sectional area than do the small nuclei but, several exceptions are recorded in the tables. In the levels of the roots farthest from the apex the increase in size in the areas measured is due chiefly to an increase in radial dimensions of the cells.

Nucleolar volumes at different levels in the root. In table 6 it can be seen that in the root of T. sprengeri the highest nucleolar volumes occur at the 459 micron level. It is at this level that the average nuclear size is the greatest. The lowest nucleolar volumes occur at the 27 micron

level where the average nuclear size is also the lowest. At the 1368 micron level the average nuclear size is slightly higher than at the 27 micron level as is also the average nucleolar volume. At the 27 micron level the mean number of nucleoli per nucleus for 101 nuclei was 4.82; at the 459 micron level it was 3.66 for 114 nuclei; at the 1368 micron level it was 3.41 for 140 nuclei counted.

The data in table 6 shows that the higher the nucleolar number the lower the total nucleolar volume. For example, at the 459 micron level the one-nucleolate nuclei have a total average nucleolar volume of 33,609.8 cubic scale units whereas the nuclei with six nucleoli at the same level have a total average nucleolar volume of only 12,879.3 cubic scale units - a reduction of over fifty percent.

In the root of Carrara the same type of determinations (table 4) were made at 423 and 891 micron levels. The nuclei at the former level average decidedly larger than at the latter level. The same relationship holds for the nucleolar volumes. At the 891 micron level the average total nucleolar volume for two-nucleolate nuclei is less than one-third the volume at the 423 micron level.

From the maximum and minimum nucleolar volumes listed in tables 4 and 6 it can be seen that while the average total nucleolar volumes show the gradations just referred to, in passing in the roots from one level to the next, the observed minimum and maximum total nucleolar volumes for indi-

vidual nuclei do not necessarily show any such gradations. The maximum nucleolar volumes observed nearly always occur in nuclei larger than the average whereas the minimum nucleolar volumes usually occur in nuclei smaller than the average.

Volumetric measurements above the levels indicated in the tables have not been made. However, by direct inspection of serial cross sections, and serial longitudinal sections of several different tulips, it was clear that the nucleolar volumes are less in regions above the active mitotic region than in regions nearer the growing point. The largest nucleoli occur in or very near to the region of most active mitosis. It is in this region that the nuclei attain their greatest dimensions.

Nucleolar volume in relation to chromosome number.

Table 7 summarizes some measurements from diploid and heteroploid cells in a root of La Reine Maximus. Measurements from a root of Carrara (data from table 4) and from one root of the triploid Inglescombe Yellow (data from table 1) are also included. The measurements are not strictly comparable due to differences in relative levels in the roots concerned. To make them so would have required more measurements than were possible in this study. The diploid and heteroploid areas in the one root, however, offered excellent material for comparisons. The normal nuclei were measured in an area

Material Determined	Avg. cell size (1)	Avg. nuclear size (3)	Avg. total nucleolar volume (2)	Avg. nucleolar no. for nuclei measured	Minimum nucleolar volume observed	Maximum nucleolar volume observed
24 chromosome cells in La Reine Maximus.	5.79	1.15	20,903.9	4.8	14,023.6	25,171.0
DO but 72 ? chromosome cells.	14.70	3.63	43,419.6	23.0	35,446.1	48,380.8
Carrara 24 chromosomes	4.66	1.70	35,941.2	4.0	24,844.5	50,507.1
Inglescombe yellow 36 chromosomes	not det.	1.70	32,160.0	6.5	21,075.6	40,879.6

Table 7. Nucleolar volume in relation to chromosome number.

- (1) Nuclear size in square inches X 2000;
(2) Nucleolar volume in cubic scale units.

immediately surrounding the chimeral region. In the case of Carrara the data was obtained from an area likely to have relatively higher nucleolar volumes than the region used in Inglescombe yellow. The data in table 7 shows that in increase in chromosome number is accompanied by an increase in total nucleolar volume in the case of the root of La Reine maximus but, that there is little if any significant difference between the diploid Carrara and the triploid Inglescombe yellow. The former even had a slightly higher nucleolar volume. These two tulips were selected because of their similarity. To make accurate comparisons of this type it is obvious that all of the clons should be of the same genetic origin i.e., absolute auto-polyploids derived by some means of vegetative reproduction. Such tulips of absolutely certain origin, were not available.

Pre-Meiotic stages

Material has been very limited for studies of pre-meiotic stages. The data here presented was obtained from longitudinal sections of anthers. It was difficult to get a fixation and stain that differentiated the nucleoli clearly. This portion of the investigation was undertaken to determine the nucleolar changes, if any, that occur between the last pre-meiotic interphase and the heterotype prophase.

Nucleolar number. Nucleolar counts have been made in three tulips, i.e., the two diploids Mrs. Moon and John

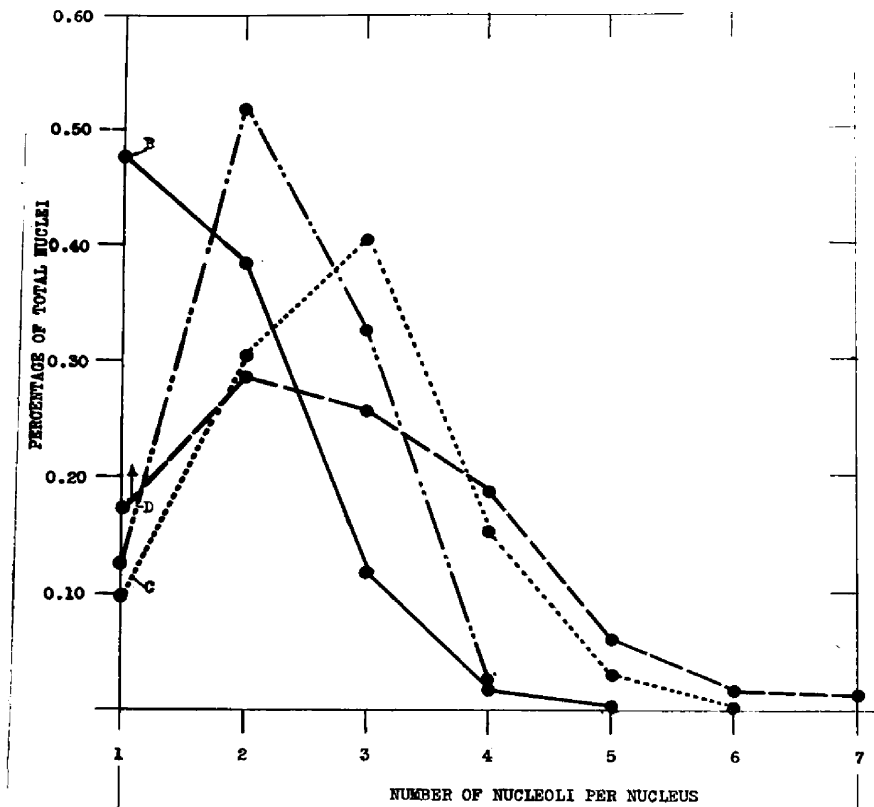


Figure 13. - Nucleolar frequencies in pre-meiotic interphases and first meiotic prophase in anthers of Mrs. Moon and John Ruskin: A, Mrs. Moon, pre-meiotic interphase, divisions still occurring, from 263 nuclei with an average nucleolar number of 2.77; B, Mrs. Moon, first meiotic prophase, 105 nuclei with average nucleolar number of 1.70; C, John Ruskin, last pre-meiotic interphase, from 507 nuclei with an average nucleolar number of 2.72; D, John Ruskin, first meiotic prophase, from 224 nuclei with an average nucleolar number of 2.25.

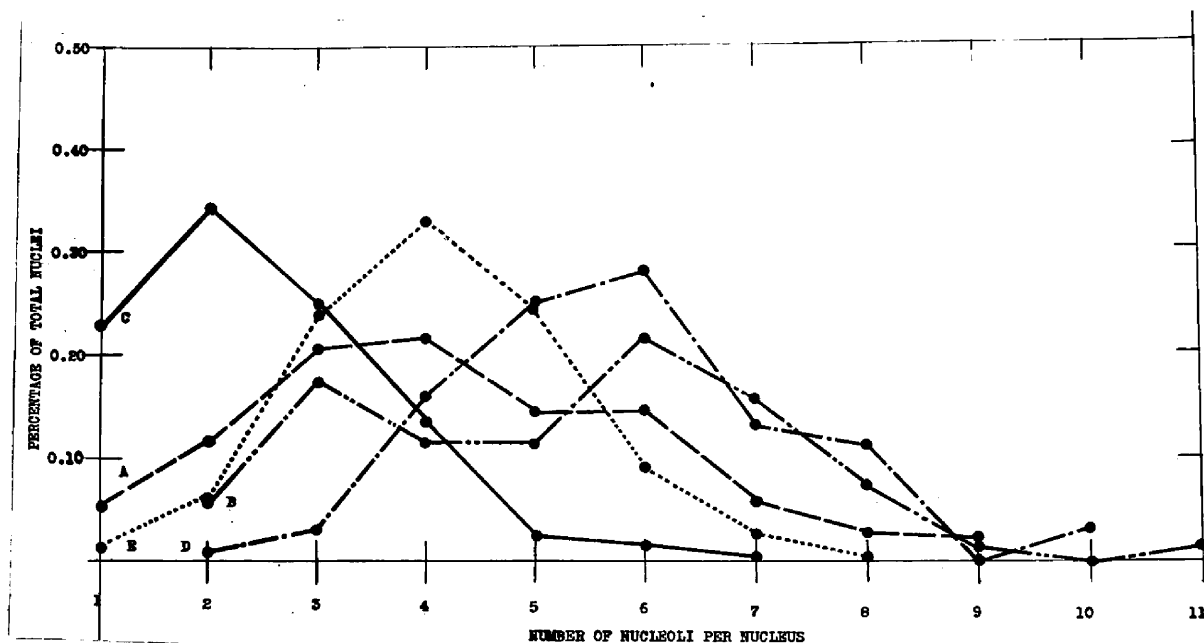


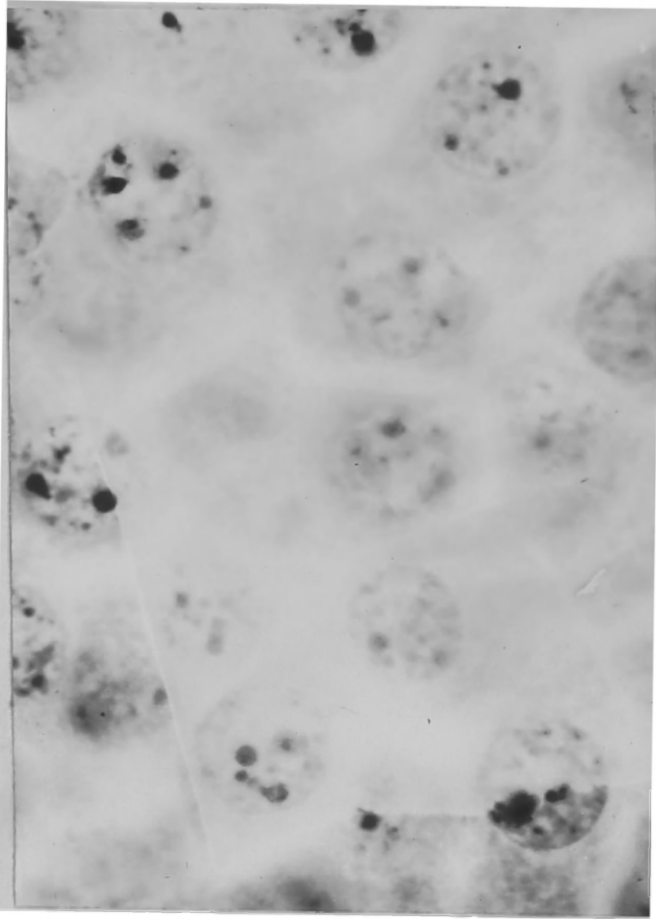
Figure 14. - Nucleolar frequencies in pre-meiotic and meiotic stages in anthers of the triploid Inglescombe Yellow:

- A, Pre-meiotic interphase, divisions still occurring in anther, from 207 nuclei with an average nucleolar number of 4.25;
- B, last pre-meiotic interphase, from 69 nuclei with an average nucleolar number of 5.02;
- C, first meiotic prophase, from 204 nuclei with an average nucleolar number of 2.44;
- D, dyad nuclei, from 100 nuclei with an average nucleolar number of 5.77;
- E, microspores in tetrad stage, from 400 nuclei with an average nucleolar number of 4.13.

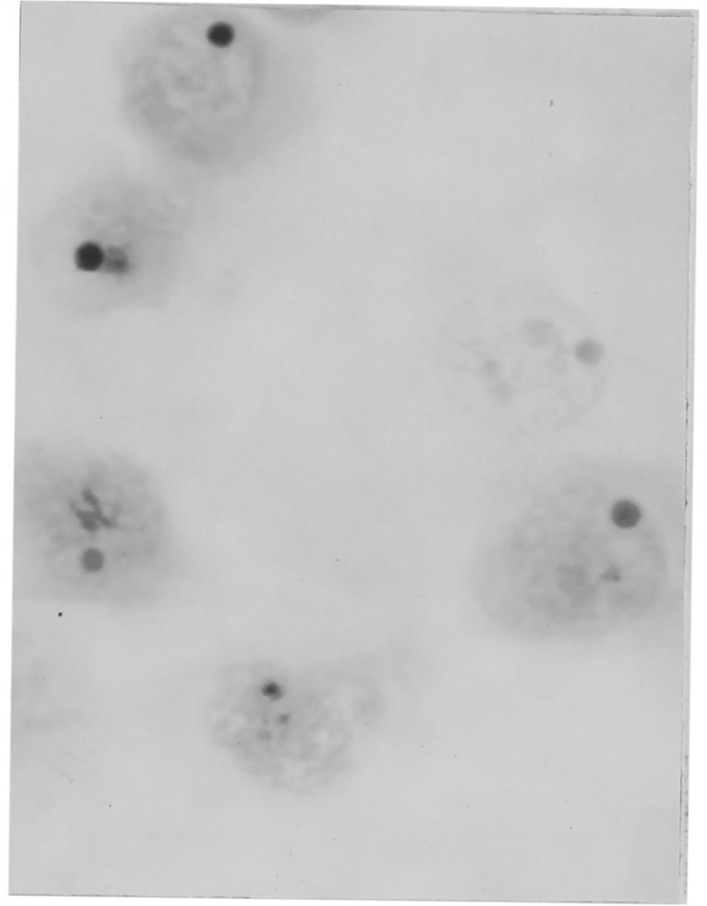
(Note: small nucleolar granules in dyad and tetrad nuclei not counted. See text)

Ruskin and the triploid Inglescombe yellow. In figure 13 the nucleolar numbers per nucleus have been plotted against their frequencies for pre-meiotic interphases and the first meiotic prophase stages in the two diploids. The former stages observed in Mrs. Moon represent a condition found several weeks before the advent of the meiotic prophase. At this time nuclear divisions were still frequent in the microsporogenous tissue. The same stages for Inglescombe Yellow are represented graphically in figure 14.

In the pre-meiotic stages in Mrs. Moon most of the nuclei possessed one large nucleolus with several small additional ones. Nuclei with five, or less, nearly equal-sized nucleoli, however, were fairly common. The graphs do not take these size differences into account. The mean number of nucleoli per nucleus at this stage in Mrs. Moon was approximately 2.8, whereas the first meiotic prophase was characterized by a mean nucleolar number of about 1.7. Nuclei with seven nucleoli were found at the former stage but, none with more than five at the latter. The pre-meiotic nucleoli were typically much more uniformly spherical in shape than root-tip nucleoli. They also lacked the distinct and large "bud"-like structure so characteristic of the first meiotic prophase in all tulips examined. Somewhat similar, though smaller, structures have been observed on pre-meiotic nucleoli. These are probably satellites.



A



B

Figure 15. - Inglescombe Yellow: A, Pollen mother cells in last pre-meiotic interphase; B, pollen mother cells in first meiotic prophase. Photomicrographs X ap. 600.

Material was obtained from John Ruskin and Inglescombe Yellow that unquestionably represented the last pre-meiotic interphase. It was collected in September about ten days before the heterotype prophase was observed in any bulbs of these varieties. The nuclei in the last pre-meiotic interphase were all very uniform in size and appearance (fig. 15,A). All of the nuclei in the sporogenous tracts were in the interphase condition. Some anthers of John Ruskin, collected just before the cessation of divisions were very similar in appearance except for slightly smaller nuclei and the presence of a few nuclei still in the process of division. The bud-like formations typical of the meiotic prophase were not observed on pre-meiotic nucleoli of John Ruskin but, small stalked spheres resembling satellites were sometimes found attached to nucleoli of Inglescombe Yellow at these stages. In both tulips the average and maximum numbers of nucleoli were higher in the interphase preceding the heterotype prophase than at the latter stage (figs. 13,14,15).

Meiotic Stages

The nucleolar numbers and volumes remain fairly constant from leptonema to late diakinesis. The nucleolar counts and measurements here reported were made chiefly from late zygonema or early pachynema to early diakinesis. The nucleoli and the relation of the chromosomes to them were

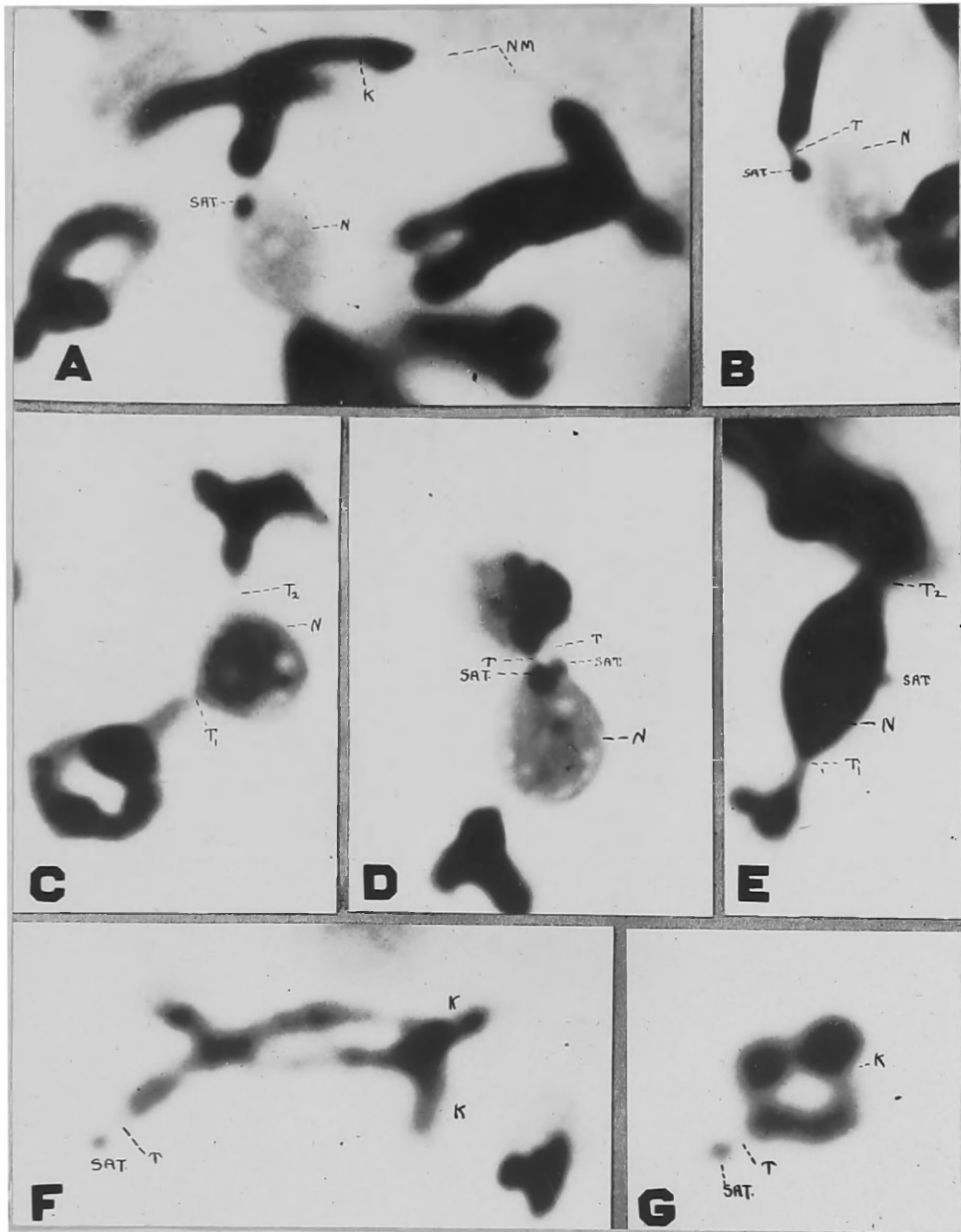


Figure 17. - Nucleoli and satellited chromosomes at diakinesis: (In all the figures SAT refers to the satellite or "bud" (see text), N refers to the nucleolus, T denotes the satellite attachment thread, K marks the position of the kinetochore and in A the nucleolar membrane is labeled NM) A, showing attachment to the nucleolus of one of the chromosome pairs heteromorphic for satellites; B, another nucleus showing same type of attachment as in A except for distinct connecting thread (T); C, nucleolus attached to two heteromorphic pairs of chromosomes; D, showing attachment of both satellites of homomorphic chromosome pair to nucleolus; E, nucleolus attached to two heteromorphic pairs of chromosomes; F, one of heteromorphic chromosome pairs showing satellite; G, homomorphic chromosome pair, satellites paired. All from the variety Adonis. Photo. X ap. 4000.

most readily studied at these stages. The first meiotic prophase was remarkably uniform in appearance with respect to nucleoli. In the diploids each pollen mother cell nucleus generally possessed one or two large nucleoli, and one or more additional small ones. Rarely three nearly equal-sized ones occurred. The mean numbers of nucleoli per pollen mother cell nucleus in the forms counted lay between 1.5 and 2.5. The numerical characteristics of meiotic nucleoli are discussed below.

Relation of the chromosomes to the nucleoli. It was not possible to accurately trace the nucleolar-chromosome relationships during the contraction. During late zygonema and pachynema, however, it was found that each nucleolus was attached to at least one pair of chromosomes. A fairly typical situation is illustrated in figure 12, plate 1. Three chromosomes were attached to the largest nucleolus and one to the smaller. There were only two nucleoli present in this nucleus. Satellites or "buds" were not visible.

Whereas all, or practically all, of the chromosomes in root-tip nuclei are attached directly to nucleoli in the prophase only a few are so attached in the meiotic prophase from leptonema to late diakinesis. The nucleoli disappear at the latter stage. In the somatic prophase the chromosomes may be attached terminally or laterally to the nucleoli (fig. 22, pl. 1). In first meiotic prophase, however, the chromosomes are apparently always attached terminally. The nucleoli during

these stages are generally characterized by a peculiar bud-like structure situated on the surface of the nucleolus (figs. 3, 9 and 11, pl. 1 also fig. 16, B, F and J). Rarely a very finely stalked small satellite-like structure was seen projecting from the "bud". As the result of a study of many nuclei from early diplotene to late diakinesis it was concluded that the nucleolar "bud" represents the place of attachment, or the approximate place of attachment of the satellites to the nucleolus. This view is in keeping with the occasional observance of smaller satellite-like structures projecting from or lying near the "bud". That the association between satellite and "bud" is usually extremely close, however, is evidenced by the condition found in most nucleoli (fig. 3, pl. 1). With sufficient destaining the "bud" usually appears as a partly projecting sphere with a darkly stained center, no other projections or satellite-like bodies being visible. In situations like those illustrated in figure 17, A, B, D, and figs. 1, 6 and 7, pl. 1, it is difficult to distinguish between "bud" and satellite. The chromatic connecting thread to the chromosome arm is often very distinct (fig. 17, B), the large darkly staining sphere (the "bud"?) appearing like a true satellite. In a few cases a second, smaller darkly staining body, was observed close to the larger sphere but the chromatic connecting thread occurred between the larger body and the chromosome arm. The different aspects of nucleolar structure

observed during the first prophase are illustrated in figure 17 and figures 1 to 12, plate 1. It seems likely that the "bud" represents a nucleolar structure with which the satellite or satellites are very closely associated or even fused. However, it also seems likely that in some cases the bud-like differentiation is absent in which case typical satellites alone are located on the region the "bud" would have occupied (fig. 1, pl. 1 and fig. 17, D). That the chromosomes themselves are firmly attached to the nucleoli is shown by the conditions illustrated in figures 4 and 9, plate 1 and, figure 17, E. The nucleolar matter must be considerably plastic to account for the spindle-shaped form frequently observed. In such cases the nucleolus and attached chromosomes look as though they had been under tension at the time of fixation.

In the single late tulip, *Adonis*, exceptionally good material was obtained for a study of satellites. In this tulip it was possible to accurately determine the number of satellited chromosomes. By observing a large number of nuclei at diakinesis it was found that there are six, and only six, of these. These are unquestionably the chromosomes that are associated directly with the nucleoli. There are four pairs of chromosomes in which one, and only one, homologue of each pair bears a satellite (figs. 5 and 8, plate 1 and fig. 17, F). There is a fifth pair in which each homologue bears a satellite. In this latter pair the satellites are usually, though not

always, closely paired in the first meiotic prophase (figs. 1 and 2, pl. 1 and fig. 17, D and G). All of the satellite-bearing chromosomes are not necessarily associated with nucleoli during diakinesis or are all of the same ones always associated with these structures. (figs. 10 and 11, pl. 1).

The satellites occur on the ends of the distal (long) arms of the chromosomes. In some cases they have been observed clearly on metaphase chromosomes in the equatorial plate stage. Two typical diakinetid nucleoli are illustrated in figures 10 and 11, plate 1. In each nucleus 12 pairs of chromosomes can be readily counted. These have been numbered more or less arbitrarily as it has not been possible to accurately distinguish all of the different morphological types characteristic of the tulip karyotype. The satellite-bearing chromosomes have also been given the same numbers in each nucleus, but the only pair which can be definitely determined is the pair morphologically homologous for satellites (No. 7).

In the nucleus shown in figure 10, plate 1, the pair morphologically homologous for satellites (number 7) is not attached to any nucleolus. Two satellite attachment threads were apparently present, but the satellites (marked S in the figure) themselves were very closely paired. Chromosome pair 12 is attached to a small spherical nucleolus (N). The large spindle-shaped nucleolus is attached to one homologue of each of two pairs of chromosomes (5 and 11). Chromo-

some pair 4 is not attached to a nucleolus but the satellite (S) on one of the homologues is distinctly visible. In figure 11, plate 1, a very different situation obtains. The chromosome pair consisting of two satellite-bearing chromosomes is here attached by one member to the principal nucleolus (note distinct "bud") and, by the other homologue to a much smaller nucleolus. These nucleoli are each attached to pairs of chromosomes morphologically non-homologous for satellites. The small nucleolus is attached to chromosome number 5 and the large one to number 4. The other two satellite-bearing chromosomes, numbers 11 and 12, are each attached to a single nucleolus. Apparently all possible combinations of association of the satellite-bearing chromosomes with the nucleoli may occur. The pollen mother cells of *Adonis* most often contained but one nucleolus (fig. 18).

In the triploid, *Inglescombe Yellow*, trivalent chromosome groups have been observed at diakinesis in which each homologue possessed a satellite. Others were apparently non-homologous with respect to satellites.

Definite connecting strands, similar to those observed in root-tip mitosis, have been observed at zygotene and diplotene of the first meiotic prophase. These may occur both laterally and terminally between the chromosomes. The inter-connections or anastomoses are apparently reduced in number as the diplonema stages move into diakinesis.

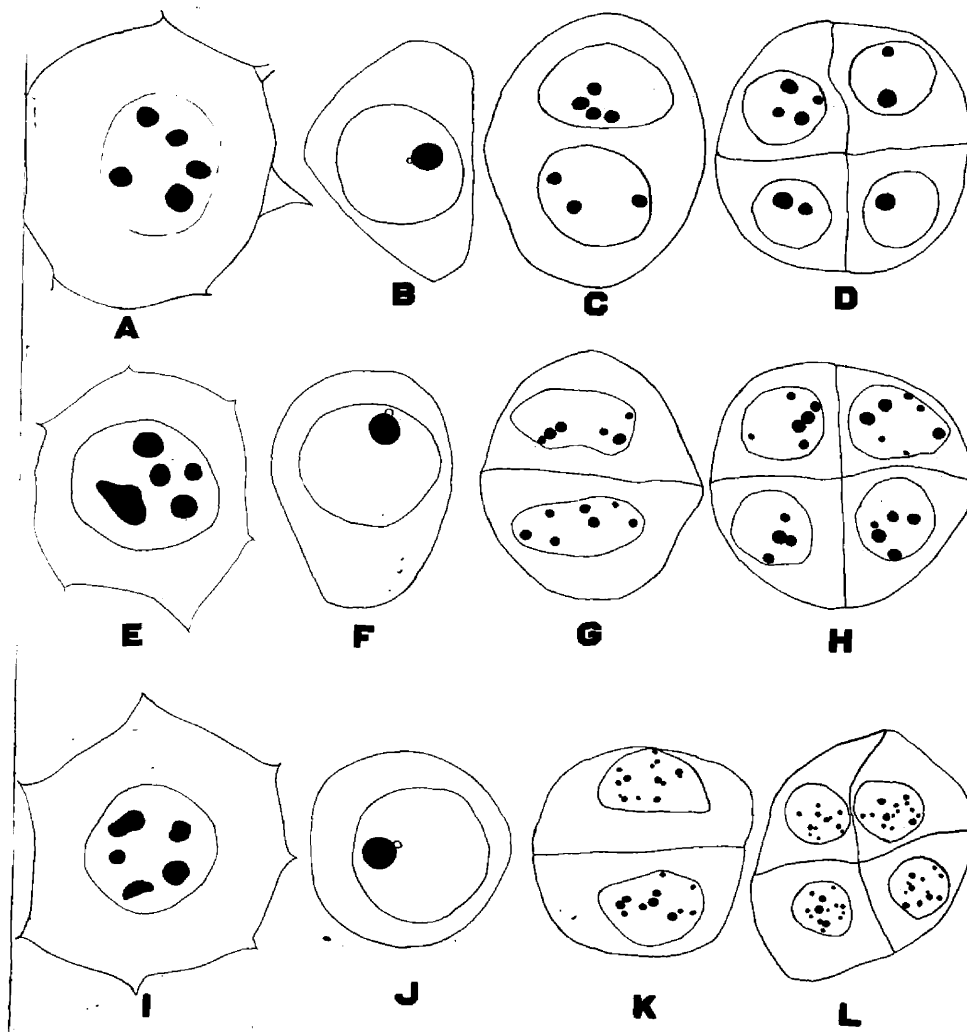


Figure 16. - Types of nucleolar organization in diploid and haploid stages of several tulip varieties: A to D, Adonis; E to H, Herman Waller; I to L, Mrs. Moon. All nuclei in interphase condition. Five, and single nucleolate nuclei were arbitrarily selected to represent root-tip and first meiotic stages. In each case the stages are arranged in the following order from left to right: root-tip, first meiotic prophase, dyad stage, and tetrad stage. Camera lucida drawings X 2000, reduced abt. $\frac{1}{2}$.

Whereas the nucleolar matter disappears in late diakinesis or early metaphase, the satellites may still be visible in the equatorial plate stage. Usually, however, they too cannot be seen after diakinesis.

Numerical relations. All of the tulips examined are characterized by remarkably low nucleolar frequencies during the first meiotic prophase. The observed frequencies are represented graphically in figures 13, 14, 18, 19 and 20. The frequencies at the second interphase or tetrad stage are also given in these figures together with the root-tip frequency for comparison.

Whereas all of the tulips examined were very similar in the first meiotic prophase striking differences occurred in the haploid or reduced phases. The three principal types observed are illustrated semi-diagrammatically by camera lucida drawings in figure 16. The first vertical column on the left, A, E and I, show typical root-tip nuclei. Five-nucleolate nuclei were arbitrarily selected. The second column, B, F and J, illustrates typical nuclei of the first meiotic prophase. Single-nucleolate nuclei were also arbitrarily chosen. The differences in nucleolar behavior responsible for this classification are illustrated in the remaining two columns.

In the first type of nucleolar organization (C and D) the number of nucleoli at first interphase and in the

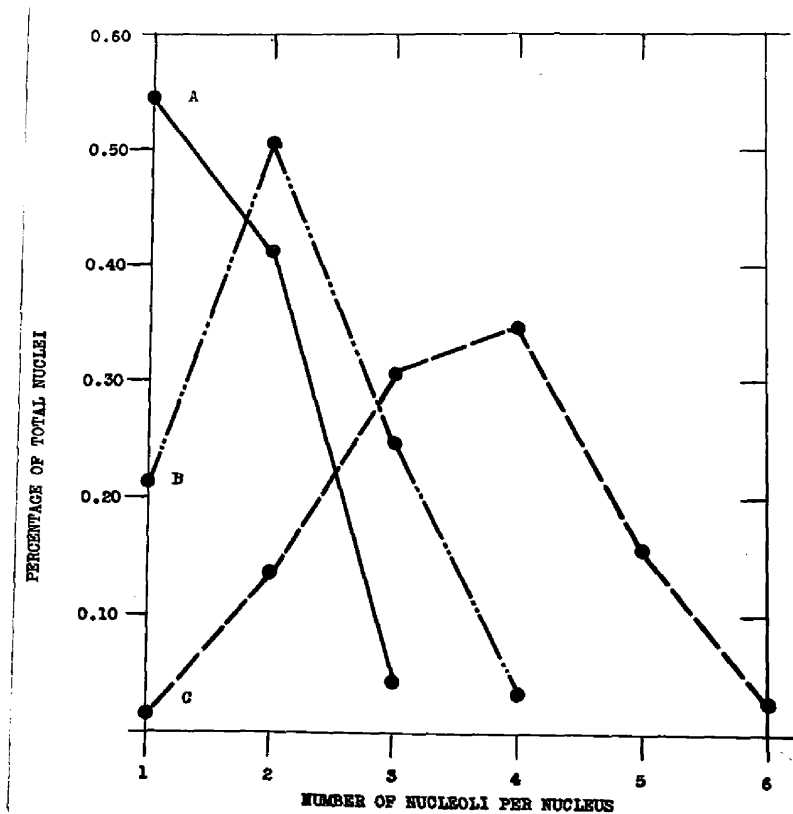


Figure 18. - Nucleolar frequencies in Adonis:
 A, first meiotic prophase, 400 pollen mother cell nuclei with average nucleolar number of 1.49;
 B, microspores in tetrad stage, 400 nuclei with average nucleolar number of 2.10;
 C, root-tip, 1000 nuclei with average nucleolar number of 3.57.

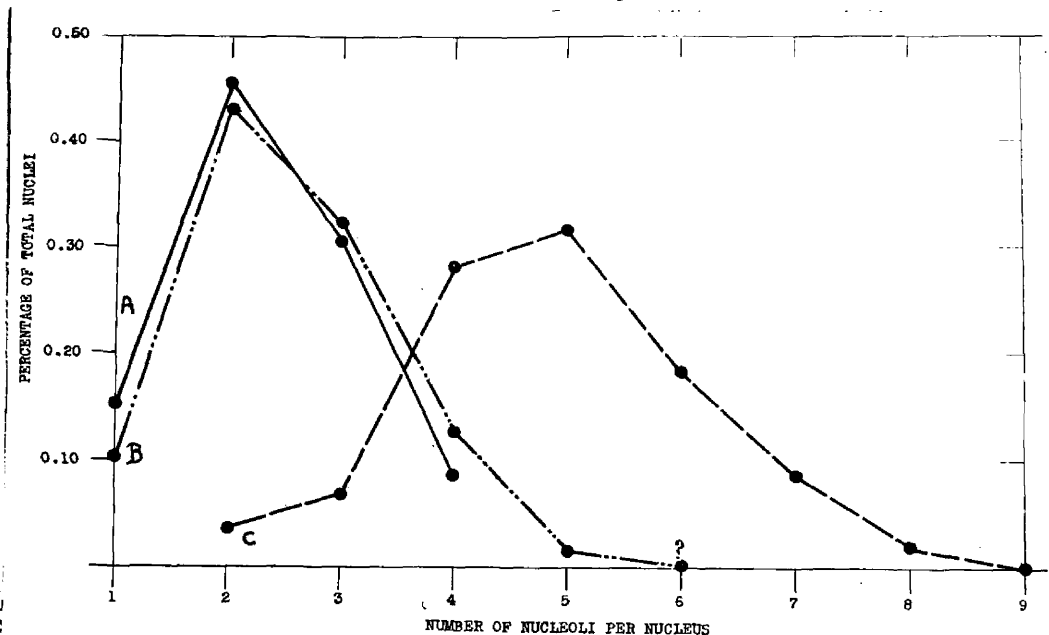


Figure 19. - Nucleolar frequencies in Gipsy:
 A, first meiotic prophase, 203 pollen mother cell nuclei with average nucleolar number of 2.33;
 B, microspores in tetrad stage, 400 nuclei with average nucleolar number of 2.53;
 C, root-tip, 500 nuclei with average nucleolar number of 4.90.

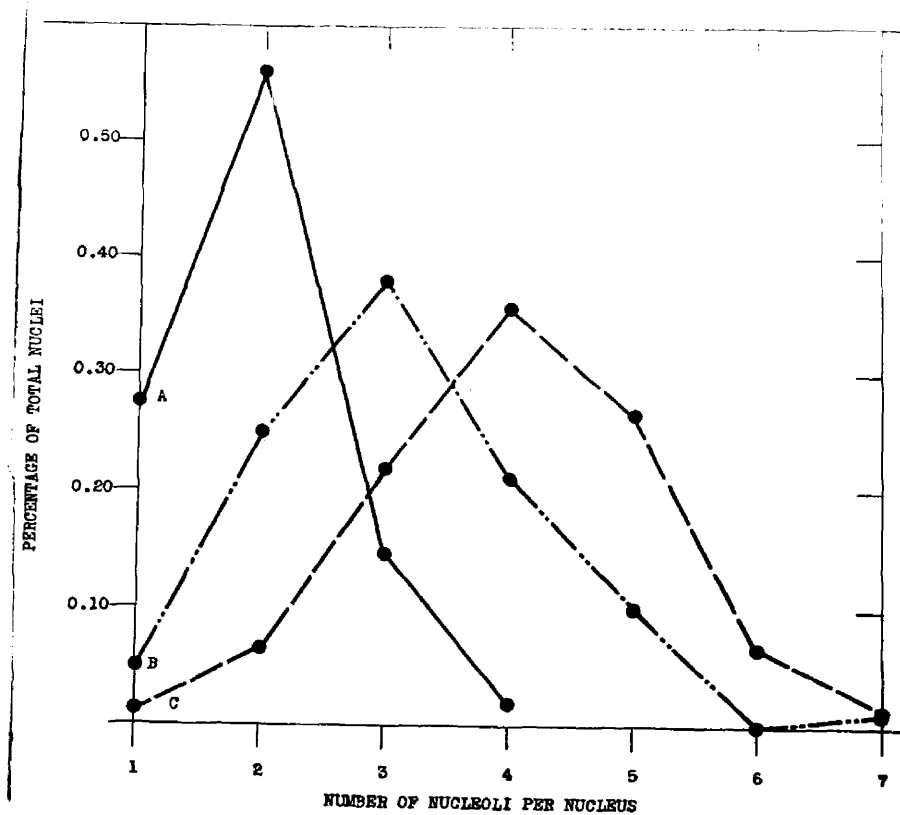


Figure 20. - Nucleolar frequencies in Herman Waller:
 A, first meiotic prophase, 200 pollen mother cell nuclei with average nucleolar number of 1.91;
 B, microspores in tetrad stage, 100 nuclei with average nucleolar number of 3.10;
 C, root-tip, 1231 nuclei with average nucleolar number of 4.04.
 (Note: nucleolar granules in microspores not counted.
 See text)

tetrad stage is always, or nearly always, fixed within definite limits. The nucleolar numbers occurring in the haploid nuclei nearly always fall within the limits possible on the theory that the maximum number of nucleoli for the root-tip is equal to the number of nucleolus-collecting or forming chromosomes. There occurs, therefore, a characteristic maximum nucleolar number for haploid nuclei. The dyad illustrated in figure 16 C constitutes the only observed exception in the tulip *Adonis*. There are a total of seven nucleoli present whereas the maximum number of nucleolus collecting chromosomes in the diploid soma is six. No exceptions were noted in the microspores, however.

In the second type of organization the nucleolar number is much less regular. Numbers of nucleoli are found at the dyad and tetrad stage that could not possibly occur if the nucleolar chromosomes all continued to function completely in the haploid cells. The extra nucleoli are usually small, however, and the larger nucleoli do follow a somewhat regular distribution (fig. 19).

The third type shows no definite nucleolar organization whatever in the haploid nuclei. The number of nucleolar bodies (or granules) frequently is above the number of chromosomes. All of the dyad and tetrad nuclei show the same lack of nucleolar organization i.e., there is no segregation of this factor.

Name and the somatic chromosome number (Det. in root-tip)	Root-tip "characteristic area"			Anther Last pre-meiotic inter-phase			Anther First meiotic prophase			Microspores Tetrad stage ⁽²⁾		
	Avg.	Max.	No. Det. (1)	Avg.	Max.	No. Det.	Avg.	Max.	No. Det.	Avg.	Max.	No. Det.
S.L. Adonis (24)	3.60	6	1000	not det.	not det.	-----	1.49	3	400	2.10	4	400
N.M. Herman Waller (24)	4.05	7	1231	DO	DO	-----	1.91	4	200	3.10 plus n	7 plus n	100
S.L. Mrs. Moon (24)	4.73	8	2640	DO	DO	-----	1.70	5	105	n	n	100 plus
S.L. John Ruskin (24)	4.89	8	4585	2.72	6	507	2.25	4	224	not det.	not det.	-----
M.M. Gipsy (24)	4.65	9	500	not det.	not det.	-----	2.33	4	203	2.53	5 or 6?	400
S.L. Inglecombe yellow (36)	7.59	13	1475	5.02	11	69	2.44	7	204	4.13 plus n	8 plus n	500

Table 8. Nucleolar numbers in certain garden tulips. Average and maximum numbers per nucleus are listed. Note: (1) total numbers of root tip nuclei counted to obtain maximum nucleolar number. Average values based on data of table 1. (2) symbol (n) refers to an indefinite number of nucleolar granules.

The above described types of nucleolar organization have been checked in both smear and paraffin-sectioned material. They are unquestionably characteristic of the particular clones investigated and do not represent the results of fixation, pre-treatment, etc. The single late or, cottage tulip, Adonis and the New Mendel tulip Gipsy belong in the first class whereas the New Mendel, Herman Waller and, the single late Ingelscombe Yellow belong in the second class. Mrs. Moon is the only representative of the third class found thus far. Other tulips have not yet been classified on this basis.

For the sake of comparison the nucleolar numerical characteristics at different stages in the life cycle have been brought together in table 8.

Volumetric relations. The nucleolar volumes at any given stage are more constant during the phases of meiosis in the anther than in root-tip nucleoli. Some variation was observed in average total nucleolar volumes following different fixatives and methods of preparation, however. For example in a permanent smear fixed in Flemming's medium fluid from one anther of a bulb of Inglescombe Yellow the average total nucleolar volume for ten pollen mother cell nuclei in first meiotic prophase was 11,177.9 cubic scale units. In the same type of material from another bulb fixed in Flemming's fluid and prepared by the paraffin technic the average total volume for ten nuclei was 10,581.0 cubic scale units. In the

former determination the maximum total volume observed was 13,645.2 cubic scale units and, the minimum 7,030.8 cubic units whereas in the latter the maximum was 14,576.0 cubic units and the minimum 7,257.6. In a third determination from another bulb the average total volume for the first prophase was 8,316.6 cubic scale units for 51 pollen mother cell nuclei. The maximum value obtained was 11,497.9 cubic units and the minimum 5,636.2 cubic units. Whereas the first two determinations were essentially the same the third was quite different.

Ten last pre-meiotic interphase nuclei in an anther of Inglescombe Yellow had an average total nucleolar volume of 12,337.4 cubic scale units. The highest volume was 16,961.6 cubic units and the lowest 8,625.7 cubic scale units. The average nuclear size was approximately 1.6 (square inches X 2000). Table 9 presents volumetric data for the nucleoli of three tulips in which measurements were made in both root-tips and sporogenous tracts of anthers. The data is intended to give only a very approximate idea of the relative volumes of nucleolar matter at different stages of the life cycle.

Name of Tulip and Chromosome number

Stage of life cycle	Measurements	Single late ADONIS (24)	New Mendel GIPSY (24)	Single late INGLEScombe YELLOW (36)
SOMATIC INTERPHASE (root tip)	Avg. Nuclear size (1)	1.33	1.77	1.70
	Max. Nucleolar volume (2)	32,131.9	55,016.2	40,879.6
	Avg. Nucleolar volume	18,386.4	37,819.7	32,160.0
	Min. Nucleolar volume	8,796.4	29,102.7	21,075.6
	Avg. no. Nucleoli in nuclei measured (3)	3.6	5.8	6.5
	Number of Nuclei measured	10	10	10
FIRST MEIOTIC PROPHASE (anther)	Avg. Nuclear size	1.44	1.42	2.54
	Max. Nucleolar volume	12,804.2	14,973.4	13,645.2
	Avg. Nucleolar volume	10,055.5	8,800.8	11,177.9
	Min. Nucleolar volume	6,633.8	5,662.1	7,030.8
	Avg. no. Nucleoli in nuclei measured	1.5	2.2	1.7
	Number of Nuclei measured	20	10	10
MICROSPORES IN TETRADS (anther)	Avg. Nuclear size	0.51	0.73	0.60 (a)
	Max. Nucleolar volume	2,358.3	3,175.2	4,258.5
	Avg. Nucleolar volume	1,323.2	2,092.0	2,597.7 (b)
	Min. Nucleolar volume	742.7	1,584.3	1,828.6
	Avg. no. Nucleoli in nuclei measured	2.0	2.5	3.9
	Number of Nuclei measured	40	20	a --- 52 b --- 88

Table 9. Nucleolar volumes compared in root tips, first meiotic prophase and microspores. Note: (1) avg. nuclear size expressed in cross-sectional areas as square inches X 2000; (2) volumes in cubic scale units; (3) representative nuclei arbitrarily selected for measurement. For characteristic means see Table. 1.

DISCUSSION

There can be little doubt that the nucleolar material (plastin) is developed by or on the chromosomes in telophase. The observations of many workers, Van Camp (29), Dermen (1), Sorokin (25), Zirkle (33), Wager (30), Heitz (5), McClintock (11), etc. all support this view. There has been considerable divergence of opinion, however, as to the exact method of origin. Heitz (5), for example, as a result of work on a number of monocots and dicots came to the conclusion (5 pg.779) that "Die Nukleolen entstehen an (nicht aus) bestimmten chromosomen". These chromosomes he designated as nucleolus chromosomes or "SAT Chromosomen" because they possessed trabants or satellites set off by achromatic threads or constructions the latter being "Thymonukleinsäure-frein Stellen" (i.e., "sine acido thymonucleinico"). The nucleolus, according to him, develops in this achromatic region behind the satellite. It is important to stress his use of the word "an" in contrast to "aus" to denote the origin of the nucleolus. Heitz also observed that the nucleolar number in telophase ("primarzahl") was equivalent to the number of SAT-chromosomes characteristic of the species. He (6) later showed, however, that when small extra nuclei were formed by lagging or fragmented chromosomes these might form a nucleolus even when the SAT-region was absent. From this he concluded that all chromosomes give off the nucleolar

material but that the SAT-chromosomes perform the function of organizing this into a definite number of bodies.

McClintock (12) has shown in Zea that it is not the achromatic stalk of the satellite but a specialized region just behind this that is the real nucleolus-forming region. In contrast to the above views Schaeede (20), working with Allium, Lilium, etc., came to the conclusion that the nucleoli form not from the telophase chromosomes but rather result from a condensation of small vacuole-like particles that originate in the nuclear sap. Sorokin (25) found that the nucleoli contain at least part of the actual chromatin of the chromosomes in Ranunculus, Anemonella, and Isopyrum. According to Dermen (1) the nucleolus may represent a by-product of the chromosome matrix and the nuclear sap formed as a surface phenomenon which can be explained by the assumption that a chemical reaction takes place between the surface substance of the chromosomes and the surrounding medium. Zirkle (33, pg.413) as a result of his studies on Zea concluded that "as the chromosomes unite to form the daughter nuclei, and as the nuclear membranes are formed, the nucleolar material collects into several droplets. These droplets, united by stainable threads flow together and form the nucleolus of the resting cell".

Zirkle's description for the telophase in Zea seems to fit the situation in Tulipa. In the latter genus the chromosomes establish inter- and intra-connections between their arms during anaphase and telophase. These anastomoses

appear to originate from the matrix substance and may well facilitate the aggregation of plastin in the telophase. Inter-connected masses of the latter material as described by Zirkle (33) for Zea also occur in Tulipa (fig.2 C, E, F, G). The plastin is apparently matrical in origin, originating from the mass of anastomosed telophase chromosomes. There are present in each telophase nucleus a definite number of nucleolus-organizing centers which eventually, with or without fusion of adjacent centers, become the interphase nucleoli. It is believed that these plastin-collecting centers represent the positions occupied by the nucleolus-forming regions of certain chromosomes of the complement although this cannot be observed in mitosis. However, from evidence obtained at meiotic stages it seems very probable that it is the satellite-bearing ends of certain chromosomes that so function.

Much has been written as to the relation between the prophase chromosomes and the nucleolus. The majority of workers in this field have described the definite attachment of the chromosomes to the nucleolus in somatic prophases. Zirkle (33, fig. 2) in Zea figures distinct attachment of the "spireme" to the nucleolus. His figures for Pinus (34, fig. 27, pl.6, figs. 1,2,3, etc., pl.3) are very clear and definite. Wager (30) presents illustrations showing clearly the attachment of the prophase chromosomes to the nucleolus. Such attachments to the nucleolus are figured abundantly in

both nucleolar and other cytological literature. These cover a wide range of micro-technical methods and material. There can be no doubt that the chromosomes of many organisms are regularly attached to the nucleoli in somatic prophase. There cannot be the slightest doubt of this in Tulipa. In this genus, at least, it seems possible that the actual attachment of the chromosomes to the nucleolus occurs in early telophase. This has not been substantiated, but there is an indication of nucleolar-chromonema attachments in the interphase. Zirkle (33), however, was unable to definitely demonstrate connection of the chromosomes to the nucleolus at interphase in Zea. The characteristic "bouquet" arrangement of all or most of the chromosomes (fig.1, H and 2, J,K,L) about the nucleoli suggests that some sort of an attraction may exist between the nucleolus-forming centers and the chromosomes. It is possible that the activities of nucleolus organization accomplished by the nucleolus-forming chromosomes brings this about.

Dermen (1) seems to attach little significance to the attachment of the chromosomes to the nucleolus. In fact (pg. 314) he " ---found no connection between the nucleolus and chromosomes in Yucca." As a result of applying reagents to living stigma hair cells of Callisia Dermen (1 pg.305) came to the conclusion "---- that even at early stages there is no complete direct association between all the chromosomes and the nucleolus and that a single attachment point between

a chromosome and a nucleolus can not be considered of any physiological importance".

As already mentioned, Zirkle's (34) drawings and photomicrographs of nuclei in Pinus are unusually clear and convincing with respect to the attachment of the chromosomes to the nucleolus. Dermen (1) likewise worked with Pinus but, due to technical difficulties did not figure the chromosomes in any of his illustrations of this genus in which nucleoli were involved. Two of his figures are of cells in Pinus strobus, the same species used by Zirkle. Frew and Bowen (3), however, like Dermen were unable to establish nucleolar-chromosomal connections. In regard to Dermen's (1) view that the attachment of only one chromosome to the nucleolus could have no physiological significance the apparently general occurrence of anastomoses between chromosomes (see Sharp (21, pg.135) should be considered. While these may have no physiological importance whatever, it should be noted that in Tulipa at least, they may persist in the root-tip until well into metaphase. If the chromosomes form an inter-connected plexus in the prophase it would be at least unwise to assume that the connection of one or a few members of the plexus with the nucleolus was without physiological significance to the other members.

Another feature of the prophase that has attracted much attention is the relation of the satellites to the nucleolus. S. Navashin (17,18) was the first to call atten-

tion to these structures. In Galtonia candicans he was able to show that the satellite-bearing chromosomes in somatic prophase were in contact with the nucleolus by their satellites. The work of Heitz (6) has already been referred to. McClintock (12) described the satellites borne on the members of chromosome pair VI of Zea mays although most of the evidence here is on meiotic chromosomes. These are definitely the nucleolus-forming chromosomes. Sorokin (25) observed structures ("nucleolosomes"), probably satellites, constantly associated with prophase nucleoli in Ranunculus abortivus, Anemonella thalictroides and Isopyrum biternatum. She was able to definitely establish the identity of "nucleolosomes" and true chromosomal satellites in R. chius. Smith (23) has presented a clear account of the satellites in relation to the nucleolus in Galtonia candicans. In somatic prophases he found at least one of the satellite chromosomes near the nucleolus and in many cases the satellite was in contact with it. Dermen (1) observed some relation between satellite chromosomes and nucleoli, but he does not seem to be convinced of its constancy or importance.

It is difficult to demonstrate the satellites in the somatic prophase of Tulipa. While they have been definitely observed at this stage it was very difficult to determine the exact number of chromosomes bearing them in root-tip nuclei. The large number and size of the chromosomes together with

the minuteness of the satellites (in most cases) make it practically impossible to do this. At metaphase satellites were more easily seen but, except for a few cases the nucleolar matter had all disappeared at this stage. The metaphase satellites are more distinct in T. sprengeri than in any other tulip examined. A maximum of seven satellited chromosomes was found in one metaphase plate but usually the number observed was much less. The highest observed number of nucleoli in this tulip is eight.

In addition to the fact that the satellites may be hidden beneath the chromosome arms the attachment thread is often so short that the satellite and chromosome arm seem to be fused. A similar situation has been reported by Holingshead and Babcock (7) in *Crepis*. The work of McClintock (12) and of M. Navashin (16) is undoubtedly of great importance in this connection. This will be considered in connection with a discussion of meiosis below. It is sufficient to say at this point with respect to Tulipa, that if the same chromosomes function in nucleolus formation in both the root-tip and in microsporogenesis Heitz's contention with respect to these chromosomes can be almost certainly verified, at least in the case of the single late tulip *Adonis*.

The interphase nucleoli of Tulipa have been studied principally in respect to their number in the root-tip. De Mol (13) was the first to call attention to the regular

numbers of nucleoli that characterize certain plants. For example, in diploid, triploid, and tetraploid varieties of Hyacinth he found maximum nucleolar numbers of two, three, and four respectively.

In the majority of investigated organisms the nucleolar number is relatively low. Thus, to mention a few, Sorokin (25) found one or two nucleoli in the Ranunculaceae investigated by her, Zirkle (33) and McClintock (12) report one and two (one in about 90% of the cells) in somatic cells of Zea mays, Dermen (1) found two and rarely three nucleoli in Callisia, Heitz (6) in Vicia faba with two SAT-chromosomes found two, and Frew and Bowen (3) a single nucleolus in Cucurbita pepo and C. maxima. Of particular interest, however, are those species in which the nucleolar number is relatively high. Zirkle (34) found nuclei containing five, six, or seven nucleoli of common occurrence in Pinus strobus. A limited number contained four or eight but the average was six. Detailed counts of a number of nucleoli are not presented. According to Zirkle (34, pg.93) plurality of nucleoli is of common occurrence in some genera. He lists Abies, Cedrus, Larix, Picea, Pseudotsuga, Sciadopitys and Tsuga as plants having high nucleolar numbers. He also states that other gymnosperms have regularly but two nucleoli in a nucleus, but that even these may be subject to variability. In the low number group he cites Agathis, Ginkgo, Juniperus, Sequoiia,

Taxus and Thuja. Dermen (1) found as many as 14 nucleoli in a cell of Pinus strobus. Schaeede (20) found three to five nucleoli in resting cells of Lilium martagon and, correspondingly more in telophase. In all of the investigations referred to the number of nucleolar bodies was more numerous in telophase than in interphase. Fusion of nucleoli in telophase probably accounts for this. In some cases at least it is doubtful whether the masses of nucleolar matter observed at this stage should be called nucleoli in the sense that they represent the definite morphological units generally implied by this term. In one way they represent nucleoli in the process of formation.

De Mol (13) has reported that the "complex" nucleoli present at the end of telophase may fragment to give several "simple" nucleoli. Discussion of prophase so-called nucleolar fragmentation will not be discussed here other than to say it almost certainly does not occur in Tulipa. As Dermen (1) has pointed out it is very doubtful if it is a real occurrence.

De Mol's (13) work on nucleolar number was very thorough, but the forms worked with by him possessed relatively low nucleolar numbers. So far as the writer is aware there has been no thorough numerical analysis of a plant with a high nucleolar number. A detailed statistical analysis of the data obtained in Tulipa will not be given in this paper. The most significant facts can be observed by inspection of the tables and graphs. Each clone is obviously characterized by a defi-

nite and characteristic maximum nucleolar number. The average number of nucleoli (see table 2) occurring in a given clon is usually roughly proportional to the maximum. Thus Adonis has a maximum of 6 and an average of 3.6. Hermen Waller is characterized by a maximum of 7 and an average of 4.0. Louis XIV has maximum and average nucleolar numbers of 9 and 5.2 respectively. Each of these varieties has a somatic chromosome complement of 24 chromosomes. In many of the tulips thousands of nuclei have been examined and the nucleolar numbers have never gone above the maximum characteristic for the different clons. The only logical conclusion that can be drawn is that a definite number of nucleolus-forming chromosomes are functioning in each case. It should be pointed out in this connection that while the exact number of satellited chromosomes have not been determined the number of these observed in some cases is quite high.- in T. sprengeri only one less than the maximum nucleolar number. The concept of a definite number of nucleolus-forming chromosomes is in keeping with the findings of Heitz (5), McClintock (12), De Mol (13) etc. on other genera. In one tulip, Adonis, it has been definitely shown that the maximum nucleolar number in the root-tip corresponds exactly to the number of satellited chromosomes (nucleolus chromosomes) occurring in the first meiotic prophase. The exact number of nucleolus-forming chromosomes at meiosis have not been determined for any other tulip.

A further study of the numerical data shows that the average nucleolar number is subject to variation but, if adjustments are made for different levels in the root an average number fairly characteristic of the clon can be obtained. As mentioned before, nucleolar numbers lower than the maximum are probably due to fusions between plastin masses that develop in the telophase. In this study it was noted that the average number of nucleoli sometimes showed a uniform variation between different levels of the same root (fig. 4). Spatial segregation of the nucleolus-forming chromosomes is obviously one of the important factors concerned here. As Heitz (5) has shown, when the nucleolus-organizing portions of two chromosomes lie close together in the telophase nucleus the chance of fusion between the two developing nucleoli is greatly enhanced.

Another point of interest in connection with nucleolar number is its relation to an increase in chromosome number through auto-polyploidy. De Mol (13) has claimed a direct correlation between the two. Dermen (1), however, was unable to demonstrate a consistent proportional relationship between the two in a polyploid series of Petunia. That auto-polyploidy in Tulipa may result in an increase in nucleolar number is clear from the situation in chimeral sectors of root-tips (fig. 11). It does not follow, however, that an

increase in chromosome number is always associated with an increase in nucleolar number. For example, T. stellata, T. chrysantha, and T. clusiana constitute a polyploid series (diploid, tetraploid and pentaploid respectively) the members of which are probably not specifically distinct although varying in certain minor characters. The 48-chromosome T. chrysantha has an average nucleolar number (see table 2) considerably lower than could be expected if a progressive increase in chromosome number were always accompanied by an equivalent increase in the number of nucleoli. Other similar cases can be found in table 2. It should be stressed that the species mentioned above are not auto-polyploids in the strictest sense, i.e., they have not all been derived from the same seed by purely vegetative propagation. There has been ample opportunity for the occurrence of a number of chromosomal irregularities due to segmental interchange, fragmentation, etc. Furthermore it seems likely that at least some of these forms are rather heterozygous. The phenomenon of "amphiplasty" discovered by H. Navashin (15,16) is of interest in connection with the seemingly irregular inheritance of the nucleolus-forming ability of certain chromosomes. This will be discussed in more detail under meiosis.

Volumetric studies of nucleoli and nuclei have shown that large nuclei generally have higher total nucleolar volumes than do small nuclei. This is in keeping with Dermen's (1) observations. Nuclei with a high number of nucleoli

usually have lower total nucleolar volumes than do nuclei with only a few nucleoli.

An increase in chromosome number may be associated with an increase in nucleolar volume (see heteroploid chimeras), but this relation does not necessarily hold between different diploid and triploid varieties, e.g., Carrara (2n) and Inglescombe Yellow (3n). Humphrey (9), however, has observed a consistent relation between chromosome number and nucleolar volume in tomato. There is little correlation, positive or negative, between nuclear size per se and nucleolar number. In Carrara (fig. 12) a slight positive correlation is indicated but the exceptions were of such a nature as to indicate little significance to the seeming relationship. A negative correlation is indicated in T. sprengeri but exceptions also occur. It seems that the relation between nuclear size and nucleolar number, if any, is at best an indirect one.

Nuclear size might appear to be related to nucleolar number because of the possible effect on spacing of the chromosomes. It is obvious, however, that the size of an interphase nucleus will not necessarily pre-determine the spatial arrangement of the chromosomes in the ensuing telophase. At the latter stage the chromosomes are always very closely associated (fig. 1, B.C.). It is important to note that Dermen (1) and Lutman (10) have both clearly shown that nucleolar volume is closely related to the nutrition of the

cell. Regardless of all the factors involved it seems almost certain that the spatial arrangement of the nucleolus-forming chromosomes at telophase, and the relative amounts of plastin generated at that stage, determine largely the number of nucleoli formed. Large amounts of plastin would encourage fusion of nucleoli and thus a reduction in their number.

The nucleolar situation in meiotic and pre-meiotic stages is rather different than in the root-tip. Differences occur in number, volume and structure of the nucleoli. There appears to be a reduction in nucleolar number between the last pre-meiotic interphase and some early stage of the first prophase. This apparently occurs previous to synapsis of the chromonemata. In Inglescombe Yellow, at least, there is apparently no reduction in nucleolar volume associated with this. Some sort of prophase fusion of nucleoli is therefore indicated. Dermen (1, pg.297) observed that "In pinus at early leptotene and during early diplotene, as many as nine nucleoli were found, the number decreasing as the development of the nucleolus advances but, the volume of total nucleoli not showing any decrease". The relation between the pre-meiotic and first prophase stages is therefore apparently similar in the two genera.

As the result of each meiotic division in Tulipa the nuclear and nucleolar volumes of each resulting nucleus

are progressively reduced but the total nucleolar volume for the whole tetrad of microspores is about the same as that of the pollen mother cell (table 9). Dermen (1, footnote pg. 305), found that in Callisia the nucleolar volume varied in different parts of the plant system but was constant in all cells at all stages in microsporogenous tissue, measuring about 4 microns. He reports that "--- the size may be about 4 microns", in the root-tip. This is quite different from Tulipa where the average nucleolar volume in the mitotic region of the root-tip is much higher than in the sporogenous cells.

The first meiotic prophase nucleoli are usually characterized by a peculiar bud-like structure which is associated with the attachments of the satellited chromosomes. The latter may not all be attached to the nucleolus but at least one is. This attachment is very definite (fig. 17, A to E). Association of the satellited chromosomes seems to be of wide occurrence but, reports as to its constancy in any one organism vary. Newton and Darlington (19, 3c, pl. I; 4, pl. II, etc.) figure distinct bud-like structures on nucleoli of Tulipa in first meiotic prophase. The only satellite illustrated in this paper occurs in a somatic metaphase plate of Keizerskroon (19, text fig. 1). None of these structures are mentioned. The attachment of the chromosomes to the nucleoli are not designated although in fig. 3c, pl. 1, the trivalent is apparently definitely attached to the nucleolus.

Smith (23) has shown definite attachment of the satellited chromosomes to the nucleolus in Galtonia candicans. The heterotype nucleoli were characterized by one or two "buds" the satellites of the chromosomes being associated with these in the constricted region. The buds are somewhat similar to the ones seen in Tulipa but in Galtonia the distinction between "bud" and satellite is much more clearly defined. Dermen (1) frequently observed "a bud-like growth" on the side of the meiotic nucleolus but, was unable to establish its relationship with other nuclear structures. He could not demonstrate more than occasional association between the satellited chromosomes and the nucleolus. In Tulipa, at least, the nucleolar "buds" are apparently of considerable significance being associated intimately with the satellites and attachment of the chromosomes to the nucleolus as before mentioned. Horton (8) in Triticum observed attachment of the satellites to the diakinetik nucleoli in about 20 percent of 2305 nuclei examined. McClintock (12) has clearly demonstrated regular attachment of the nucleolus-organizing chromosomes in Zea mays. No explanation is offered for the different results obtained by these workers but, in Tulipa the association between satellited chromosomes and nucleoli in first prophase seems to be of constant occurrence in the varieties examined.

Of particular significance in Tulipa is the fact that the satellited chromosomes are not always morphologically alike in the heterotypic prophase. Thus in Adonis there is

one pair in which each homologue bears a satellite and four pairs in which only one of each bears one of these structures. The satellited chromosomes are the only ones that are attached to the nucleolus. The occurrence of satellite differences in otherwise homologous chromosomes was reported by S. Navashin in Galtonia candicans. He described races in which either all large, all small, or both large and small satellites were present. The first two he called "symmetrical" races, the latter "assymetrical" races. In all of the races the satellites were typically associated with the nucleolus. Smith (23), however, was unable to show distinct races or forms in Galtonia candicans on the basis of Navashin's criteria. Smith (23) in referring to some work of Emme (2) states that the latter found a number of varieties of barley to be "--- polymorphic with regard to the presence or absence of one or both satellites of a particular pair of chromosomes". Emme called these chromosomes "Nebenchromosomen". In 44 varieties of Hordeum he found two with 2 of these peculiar chromosomes, 17 with one each and the rest without any. His figures do not give the impression that these structures are true satellites. The kinetochores of the chromosomes are not definitely shown, the "neben" bodies appearing much like a portion of the chromosome arm. The attachment threads are very coarse in some instances. That Emme (2 pg.232) did not consider these portions of the chromosome arm proper is clearly shown by

his statement "Das Fadchen ist scheinbar ein echter Faden und keine 'constriction' nach Sakamura". The relation of these chromosomes to the nucleolus was not considered. If these chromosomes in barley are of the nucleolus-generating type the situation is very similar to that observed in Tulipa.

The differences in nucleolus-organizing power, observed in the haploid nuclei of some tulip varieties, is of particular interest. The nucleolus-forming chromosomes may only partially lose this ability in dyad and microspore nuclei or there may be no nucleolar organization whatever at these stages. In the latter case (Mrs. Moon) the plastin apparently originates from all the chromosomes forming an indefinite number of nucleolar granules which may be more numerous than the chromosomes. Every microspore nucleus shows the same lack of organization i.e., there is no segregation of the factors responsible for this condition. Hybrids formed from an "organized" type and the "unorganized" variety (the latter used as pollen parent) show ordinary nucleolar behavior in the root-tip. The nucleolus-forming power of the chromosomes is therefore present in the hybrid. As the hybrids (see table 2, Louis XIV X Mrs. Moon) had nucleolar maxima of at least eight, some of the nucleolus-forming chromosomes must have come from the pollen parent, Mrs. Moon. This is substantiated by microspore studies in Louis XIV which show that the nucleolus-organizing chromosomes in this tulip are about equally distributed in meiosis. Louis XIV has a

maximum of 9 nucleoli per nucleus in the root-tip.

Two papers by McClintock (12) and M. Navashin (16) have already been referred to. They have an important bearing on the occurrence of asymmetry of satellite chromosomes in Tulipa and upon the peculiar types of haploid nucleolar organization just described. McClintock (12) found chromosome VI in Zea mays to be characterized by a deeply staining body just behind the satellite stalk that functions as a nucleolus-forming organ. As a result of X-ray treatment a chromosomal interchange resulted in which this body was split in two, each inter-changed chromosome getting a section. Whereas in ordinary haploid, diploid and triploid tissues possessing normal chromosome VI there were respectively one, two and three nucleoli, various peculiar nucleolar situations resulted in plants having one or more inter-changed chromosomes. Plants homozygous for the inter-change developed four nucleoli in their somatic telophases two being typically larger than the others. The sizes of the nucleoli were proportional to the sizes of the portions of the chromosome-forming bodies present. Plants heterozygous for the interchange developed three nucleoli in their somatic telophases, the microspores showing segregation. One of her very significant contributions was the demonstration of different "functional capacities" for the two types of inter-change chromosomes. When both types were present in the same nucleus the chromosome with the largest segment of the nucleolus-organizing body formed

a large nucleolus and the other chromosome a small one. When the latter was present alone in a nucleus it developed a typical large nucleolus. She found furthermore (12,pg.322) that "The activity of the nucleolar-organizing element is hindered by certain genomic deficiencies". When this happened many small nucleolus-like bodies developed but were never organized into definite nucleoli. According to her "These small nucleoli appear to develop from a swelling and later collection into droplets of the matrix substance of the chromosome". Creighton (see McClintock (12) pg. 321) observed a similar phenomenon in Zea when chromosome complements lacking the entire nucleolus-forming region of chromosome VI were formed in microsporogenesis. An indefinite number of nucleolar bodies were found in such microspores. Of interest in connection with the apparent disappearance of satellites, is McClintock's (12 pg.298) observation that the length of the attachment thread in chromosome VI varied greatly in different nuclei. This seemed to depend "---- upon the distance the satellite was removed from the nucleolar-organizing body in the previous prophase and also upon the stage at which the satellited chromosome is released from the nucleolus in the late prophase". This difference in satellite thread length carries over into metaphase.

McClintock's (12) results are very suggestive in connection with the work of M. Navashin (16) on Crepis.

Navashin (15) had, previously observed the apparent loss of a satellite in certain Crepis hybrids. This phenomenon he termed "amphiplasty". In his recent paper (16 pg.199) he reports that this change "--- is reversible and the satellited chromosomes recover their normal shape as soon as the pure complement is extracted from the hybrid by means of segregation". He found "-- that the presence of an incomplete haploid set of the one species is sufficient to cause changes in two satellited chromosomes of the other". This "loss" of the satellite he found was in reality due to its fusion with the arm of the chromosome. McClintock (12) suggests that this phenomenon in Crepis may be explained on the assumption of differential nucleolus-organizing powers of the satellited chromosomes of the hybrid. Thus, due to the failure of certain of these to function in nucleolus formation their satellite stalks never become elongated and the satellite cannot be differentiated from the chromosome arm. Navashin did not report nucleolar behavior in these plants.

It is possible that certain nucleolar phenomena observed in Tulipa can be explained on the basis of McClintock's contentions. The occurrence of different numbers of functioning nucleolus-forming chromosomes in the varieties and species-clons suggests that a situation similar to that occurring in Crepis may exist. The behavior in microsporogenesis shows conclusively that the ability of the nucleolus-forming chromosomes to function in some cases depends upon the combination

of chromosomes present. Thus, in Mrs. Moon both haploid sets are unable to function alone. In this respect the situation in Zea differs. McClintock (12) observed segregation in the abnormal behavior in microsporogenesis. As pointed out above, this does not occur in the variety of Tulipa. In the latter case it was therefore possible to definitely test the fertility of the microspores showing lack of nucleolar organization. There is little if any reduction in fertility due to this condition. This shows furthermore that nucleolar divisions can occur without the presence of a limited number of definitely organized nucleoli because the pollen grains of Mrs. Moon each contain several nuclei.

SUMMARY

(1) The development of nucleoli in mitosis is described. The nucleolar plastin appears to originate from the matrix in telophase. This material is organized into nucleoli about certain loci. If these loci, or collecting centers, lie close together the developing nucleoli fuse.

(2) A detailed numerical study of interphase root-tip nucleoli has shown that each clone is characterized by a fixed maximum nucleolar number. The average is subject to variation which seems to be in part due to the spatial arrangements of the nucleolus-organizing chromosomes and the relative volumes of plastin formed in telophase. Some or all of the chromenemata apparently remain attached to the nucleoli in interphase. Volumetric studies of interphase nuclei and nucleoli show that nuclear size and nucleolar volume tend to be directly proportional. Nuclei with a high number of nucleoli generally have a lower total nucleolar volume than do nuclei with only a few of these bodies. There appears to be little or no direct relationship between nuclear size and nucleolar number, however.

(3) All, or nearly all, of the chromosomes are definitely attached to the nucleoli in mitotic prophase, the chromosomes being inter-connected by lateral anastomoses. Satellited chromosomes were definitely observed, but it was not possible to determine the exact number of these in somatic

prophase or metaphase. The satellites are associated with the nucleoli in prophase.

(4) Autoheteroploidy (as observed in chimeras) results in an increase in nucleolar number and volume. Varieties forming a polyploid series of related forms (not strictly autopolyploids), however, may not show a consistent variation in nucleolar number and volume.

(5) A study of first meiotic prophase nucleoli has established the identity between satellited and nucleolar chromosomes in the single late tulip, Adonis, 4 pairs of chromosomes are hetero-morphic, and one pair homo-morphic for satellites. Such lack of morphological homology in pairs of chromosomes apparently occurs in some other varieties. In Adonis the number of satellited chromosomes is equivalent to the maximum number of nucleoli occurring in the root-tip (six). The morphological relationships between nucleoli and satellited chromosomes are described for the first meiotic prophase.

(6) The results of numerical and volumetric studies at the last pre-meiotic interphase and the first meiotic prophase are presented. At both of these stages the nucleolar number and volume is much less than in root-tips. Very little, if any, change in total nucleolar volume occurs between the last pre-meiotic interphase and the second meiotic interphase (considering total nucleolar volume of all four microspores.)

(7) The nucleolus-organizing ability of the chromosomes may be partly or entirely suppressed in haploid nuclei. Thus the ability of given chromosomes to form nucleoli is dependent in some cases upon the combination of chromosomes present in the nucleus. Complete lack of nucleolar organization does not prevent nuclear division in, or fertility of, the microspores. The nucleolus-organizing function of the chromosomes is present in root-tips of hybrids formed from a cross in which the pollen came from a variety showing no organization in haploid nuclei. In this case some of the nucleolus-forming chromosomes in the hybrid came from the pollen parent.

The writer wishes to acknowledge his indebtedness and appreciation to Dr. Ronald Bamford for his valuable suggestions, criticisms, and unfailing interest during the progress of this work.

BIBLIOGRAPHY

1. Dermen, H.
1933. ORIGIN AND BEHAVIOR OF THE NUCLEOLUS IN PLANTS.
Journal of the Arnold Arboretum 14: 282-323,illus.
2. Emme, H.
1925. BEITRAGE ZUR CYTOLOGIE DER GERSTEN. I. KARYOTYPEN
DER GERSTEN.
Zeitschr. Ind. Abst. Vererb. 37: 229-236,illus.
3. Frew, Priscilla, and Bowen, R. H.
1929. NUCLEOLAR BEHAVIOUR IN THE MITOSIS OF PLANT CELLS.
Quar. Jour. Micros. Sci. 73: 197-219, illus.
4. Guillermond, A, and Mangenot, G.
1928. REVUE GENERAL DES TRAVAUX DE CYTOLOGIE. LE NUCLEOLE.
Rev. Gen.Bot. 38: 251-262.
5. Heitz, E.
1930. LAGE, FORM UND GROSZE PFLANZLICHER NUKLEOLEN.
Planta 12: 775-844, illus.
6. -----
1931. NUKLEOLEN UND CHROMOSOMEN IN DER GATTUNG VICIA.
Planta 15: 495-505, illus.
7. Holingshead, Lillian and Babcock, E. B.
1930. CHROMOSOMES AND PHYLOGENY IN CREPIS.
Univ. of Calf. Publ. in Agric.Sci. 6: 1-53.
8. Horton, Ethel Sue
1936. STUDIES IN THE CYTOLOGY OF WHEAT AND OF A WHEAT
SPECIES HYBRID.
Amer.Jour.Bot. 23: 121-128, illus.
9. Humphrey, L. M.
1934. THE MEIOTIC DIVISIONS OF HAPLOID, DIPLOID AND
TETRAPLOID TOMATOES WITH SPECIAL REFERENCE TO THE
PROPHASE.
Cytologia 5: 278-300, illus.
10. Lutman, B. F.
1925. SENESCENCE AND REJUVENESCENCE IN THE CELLS OF THE
POTATO PLANT.
Univ. of Vermont Agric.Exp.Sta.Bull. 252, illus.
11. McClintock, Barbara
1931. CYTOLOGICAL OBSERVATIONS OF DEFICIENCIES INVOLVING
KNOWN GENES, TRANSLOCATIONS AND AN INVERSION IN
ZEA MAYS.
Agr.Exp.Sta. Univ.Mo.Res.Bull. 163, illus.

12. -----
 1934. THE RELATION OF A PARTICULAR CHROMOSOMAL ELEMENT
 TO THE DEVELOPMENT OF THE NUCLEOLI IN ZEA MAYS.
 Zeitschr.f.Zellforsch.u.Mikro.Anat. 11: 294-328,
 illus.
13. Mol, Wm. Edw. de.
 1928. NUCLEOLAR NUMBER AND SIZE IN DIPLOID, TRIPLOID AND
 ANEUPLOID HYACINTHS.
 La Cellule 38: 7-66, illus.
14. Montgomery, T. H.
 1898. COMPARATIVE CYTOLOGICAL STUDIES, WITH SPECIAL
 REGARD TO THE MORPHOLOGY OF THE NUCLEOLUS.
 Jour. Morph. 15: 266-582.
15. Navaschin, M.
 1927. UEBER DIE VERANDERUNG VON ZAHL UND FORM DER CHROMO-
 SOMEN INFOLGE DER HYBRIDISATION.
 Zeitschr.f.Zellforsch.u.Mikro.Anat. 6: 195-233,
 illus.
16. -----
 1934. CHROMOSOME ALTERATIONS CAUSED BY HYBRIDIZATION AND
 THEIR BEARING UPON CERTAIN GENERAL GENETIC PROBLEMS.
 Cytologia 5: 169-203, illus.
17. Navaschin, S.
 1912. SUR LE DIMORPHISME NUCLEAIRE DES CELLULES SOMATIQUES DE
 GALTONIA CANDICANS.
 Bull.Acad.Imp.Sci.St.Petersbourg 6: 373-385.
18. -----
 1927. ZELLKERNDIMORPHISMUS BEI GALTONIA CANDICANS DES. UND
 VERWANDTEN MONOKOTYLEN.
 Ber.Deutsch.Bot. Ges.45: 415-428, illus.
19. Newton, W.C.F. and Darlington, C.D.
 1929. MEIOSIS IN POLYPLOIDS. PART I. TRIPLOID AND PENTAPLOID
 TULIPS.
 Jour. of Genetics 21: 1-15, illus.
20. Schaede, R.
 1928. UBER DAS VERHALTEN DES NUKLEOLUS WAHREND DER
 KERNTHEILUNG.
 Protoplasma 5: 41-54, illus.
21. Sharp, L. W.
 1934. INTRODUCTION TO CYTOLOGY, THIRD EDITION.
 McGraw-Hill, New York, illus.

22. Smith, F. H.
1934. THE USE OF PICRIC ACID WITH THE GRAM STAIN IN PLANT CYTOLOGY.
Stain technology 9 no.3.
23. -----
1933. THE RELATION OF THE SATELLITES TO THE NUCLEOLUS IN GALTONIA CANDICANS.
Amer.Jour. of Bot. 20: 188-195, illus.
24. Smith, Gilbert M.
1933. THE FRESH-WATER ALGAE OF THE UNITED STATES, FIRST EDITION.
McGraw-Hill, New York, illus.
25. Sorokin, Helen
1929. IDIOGRAMS, NUCLEOLI, AND SATELLITES OF CERTAIN RANUNCULACEAE.
Amer. Jour.Bot. 16: 407-420, illus.
26. Tanner, Fred. W.
1928. BACTERIOLOGY
John Wiley & Sons, New York, illus.
27. Taylor, W. R.
1926. CHROMOSOME MORPHOLOGY IN FRITILLARIA ALSTROEMERIA, SILPHIUM, AND OTHER GENERA.
Amer.Jour.Bot. 13: 179-193, illus.
28. -----
1924. THE SMEAR METHOD FOR PLANT CYTOLOGY.
Botanical Gazette 78: 236-278.
29. Van Camp, G. M.
1924. LE ROLE DU NUCLEOLE DANS LA CARYOCINESE SOMATIQUE.
La Cellule 43: 7-50, illus.
30. Wager, H.
1904. THE NUCLEOLUS AND NUCLEAR DIVISION IN THE ROOT APEX OF PHASEOLUS.
Ann.Bot. 18: 29-55, 1904.
31. Wilson, E.B.
1925. THE CELL IN DEVELOPMENT AND HEREDITY.
MacMillan, New York, illus.
32. Yamaha, G. and Sinoto, Y.
1925. ON THE BEHAVIOR OF THE NUCLEOLUS IN THE SOMATIC MITOSIS OF HIGHER PLANTS, WITH MICROCHEMICAL NOTES.
Bot.Mag.Tokyo. 39: 205-219, illus.
33. Zirkle, Conway
1928. NUCLEOLUS IN ROOT TIP MITOSIS IN ZEA MAYS.
Bot. Gaz. 86: 402-418, illus.
34. -----
1931. NUCLEOLI OF THE ROOT TIP AND CAMBIUM OF PINUS STROBUS.
Cytologia 2: 7-105, illus.

Explanation of plate 1.

(Figs. 1-11, diakinesis stages in single last tulip *Adonis*.
Flemming's fixation)

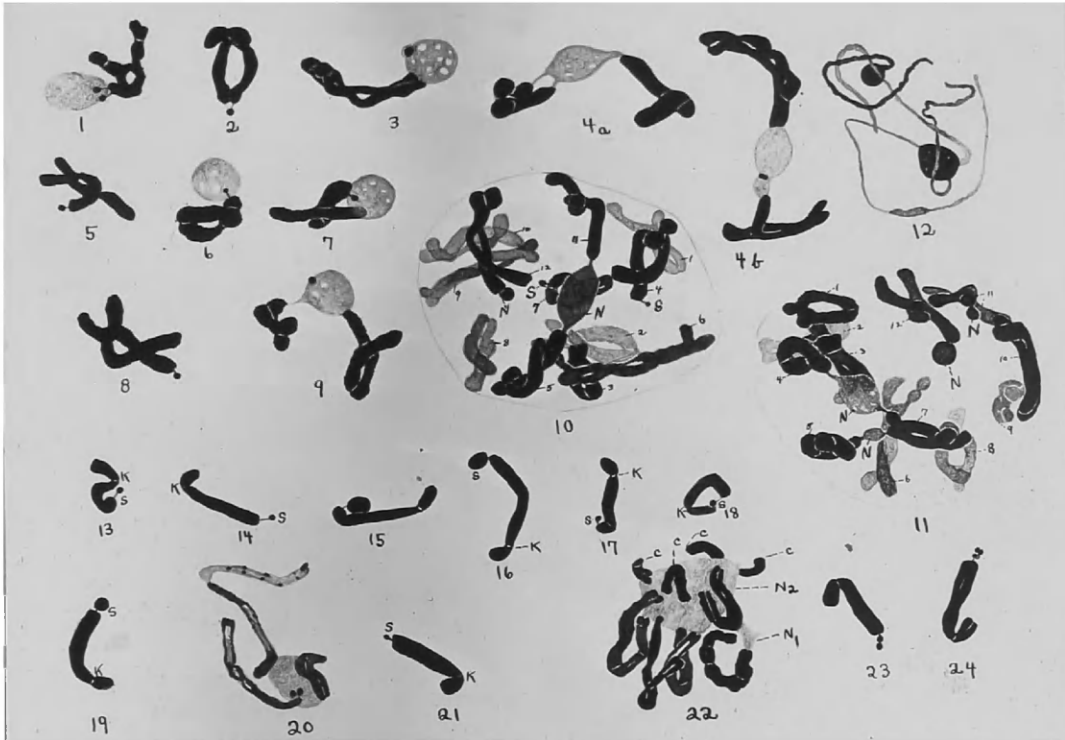
- Fig. 1. Homomorphic pair of chromosomes attached to nucleolus (same shown in photomicrograph, fig.17, D).
Fig. 2. Homomorphic pair of chromosomes, satellites closely paired.
Fig. 3. Homomorphic pair of chromosomes attached to nucleolus, satellites cannot be distinguished from the "bud".
Fig. 4a. Nucleolus attached to homomorphic, and one pair of heteromorphic chromosomes.
Fig. 4b. Nucleolus attached to two of the heteromorphic pairs of chromosomes.
Fig. 5. One of the heteromorphic pairs of chromosomes.
Figs. 6 & 7. Nucleoli attached to heteromorphic pairs of chromosomes.
Fig. 8. Heteromorphic pair of chromosomes.
Fig. 9. Nucleolus attached to two pairs of heteromorphic chromosomes.
Figs. 10 & 11. Two diakinetik nuclei each containing 12 bivalents. Nucleoli labeled (N), satellites (S), the nucleolus chromosomes are numbered the same in both nuclei, i.e., 4,5,7,11,12. Pair 7 is homomorphic for satellites. (See text for complete description).
Figs. 12. Two nucleoli with attached chromosomes from a nucleus in pachytene.
Figs. 13 to 18. Satellited chromosomes from root-tip metaphase plates of *Adonis*. Remnants of nucleolar matter are still attached in 15 and 16.

(Figs. 19 to 22 from root-tip cells of *T.eichleri*. Bouin's fixation.)

- Fig. 19. Satellited chromosome at metaphase with associated nucleolar material.
Fig. 20. Satellited and non-satellited chromosomes attached to prophase nucleolus.
Fig. 21. Satellited chromosome at metaphase.
Fig. 22. Nucleolus with associated chromosomes at late prophase. Nucleolar matter (plastin) is marked (N). Chromosomes marked (c)-have been cut transversely in sectioning.
Figs. 23 and 24. Tandem satellited chromosome of the Darwin, Allard Pierson. The satellites are divided in 24.

Camera lucida drawings X 2000. Reduced abt. 1/8.

Plate 1



Nucleoli in Tulipa

APPROVED Ronald Bamford

May 16, 1936